

*Protein Network  
Comparative  
Genomics*

**Trey Ideker**

**University of California San Diego**

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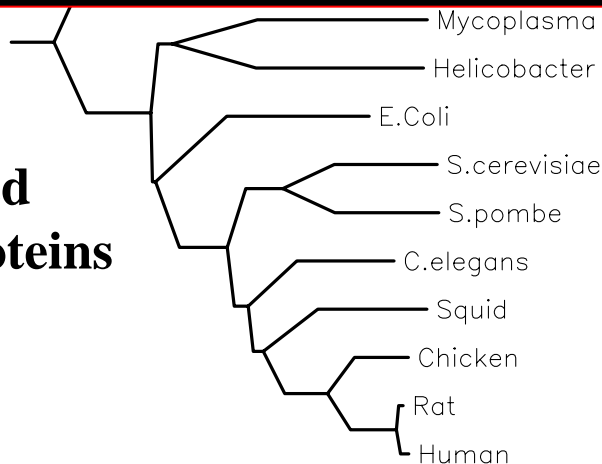
# From protein sequences... to protein networks

Query Sequence  
GACTGCATTAC

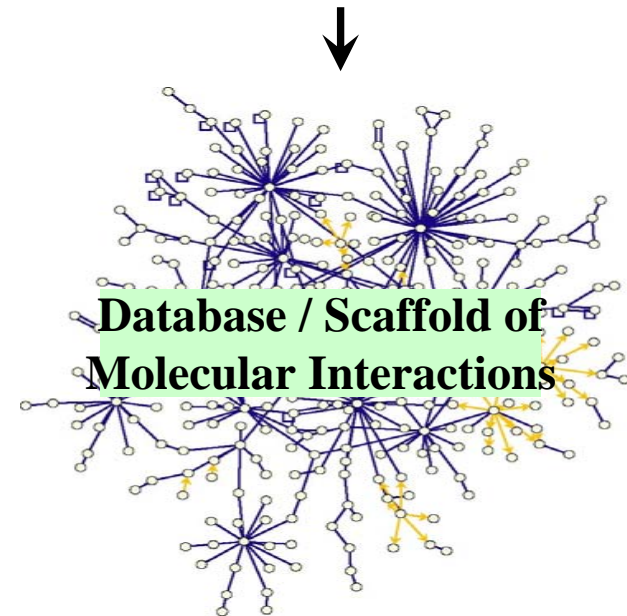
**Two goals:**

- 1) *Screen out false positives and non-functional interactions*
- 2) *Place into pathway models*

**Conserved genes/proteins**



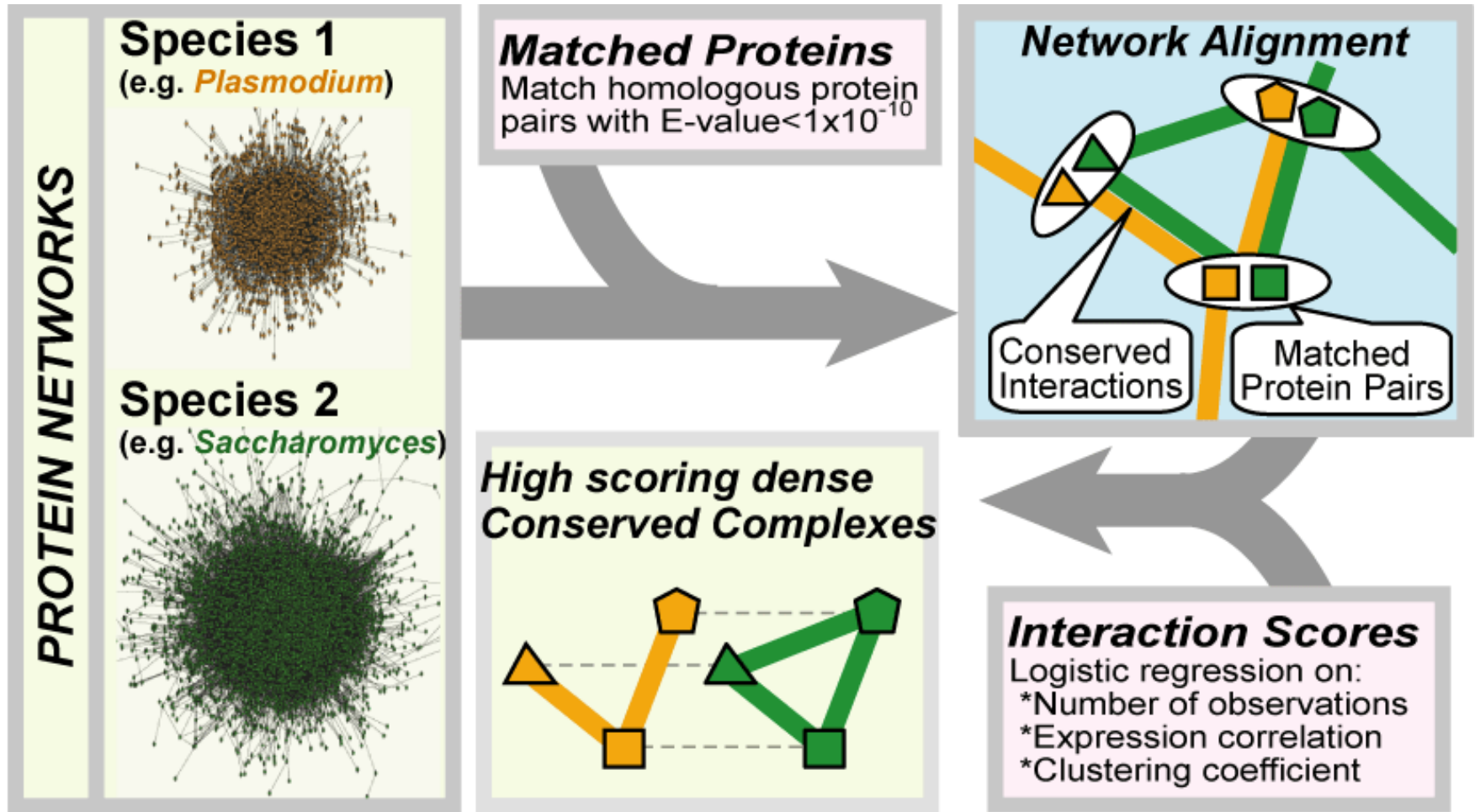
Global query



**Interaction pathway and complexes associated with query data**

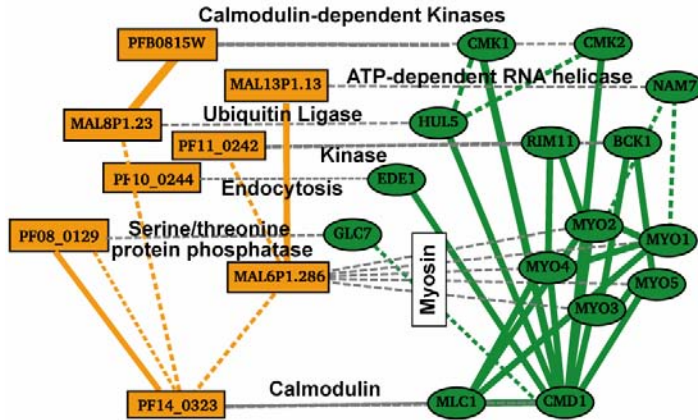
# Cross-comparison of networks:

- (1) Conserved regions in the presence vs. absence of stimulus
- (2) Conserved regions across different species

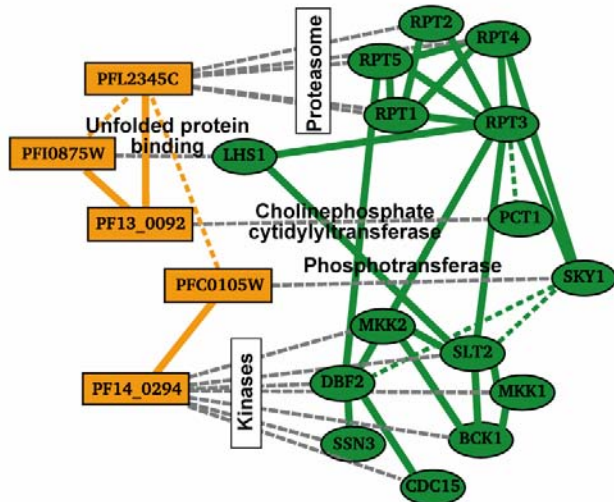


# Plasmodium: a network apart?

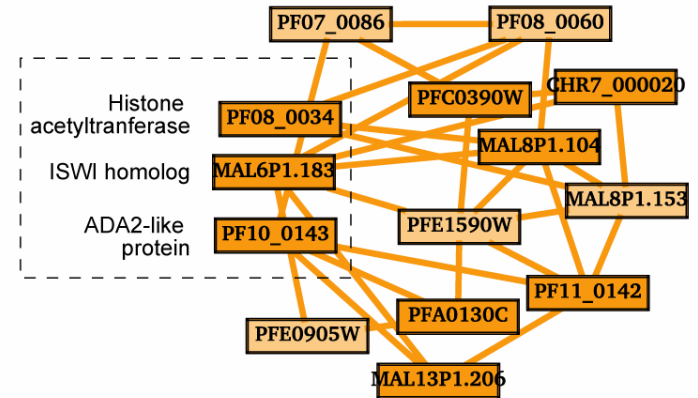
[a] Endocytosis



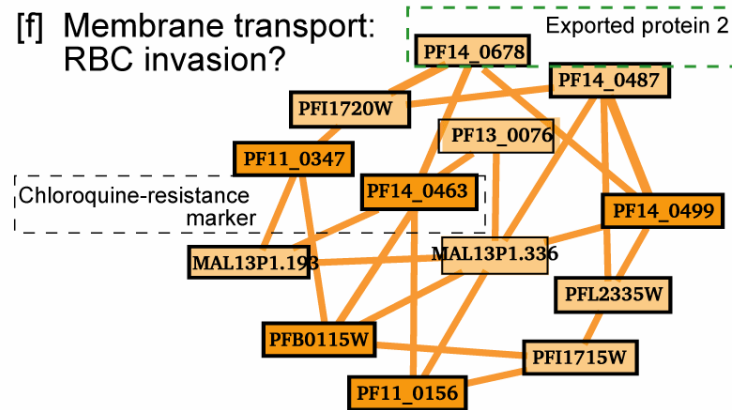
[b] Unfolded protein response



[e] Chromatin remodeling



[f] Membrane transport: RBC invasion?



*Plasmodium*-specific protein complexes

Conserved *Plasmodium* / *Saccharomyces* protein complexes

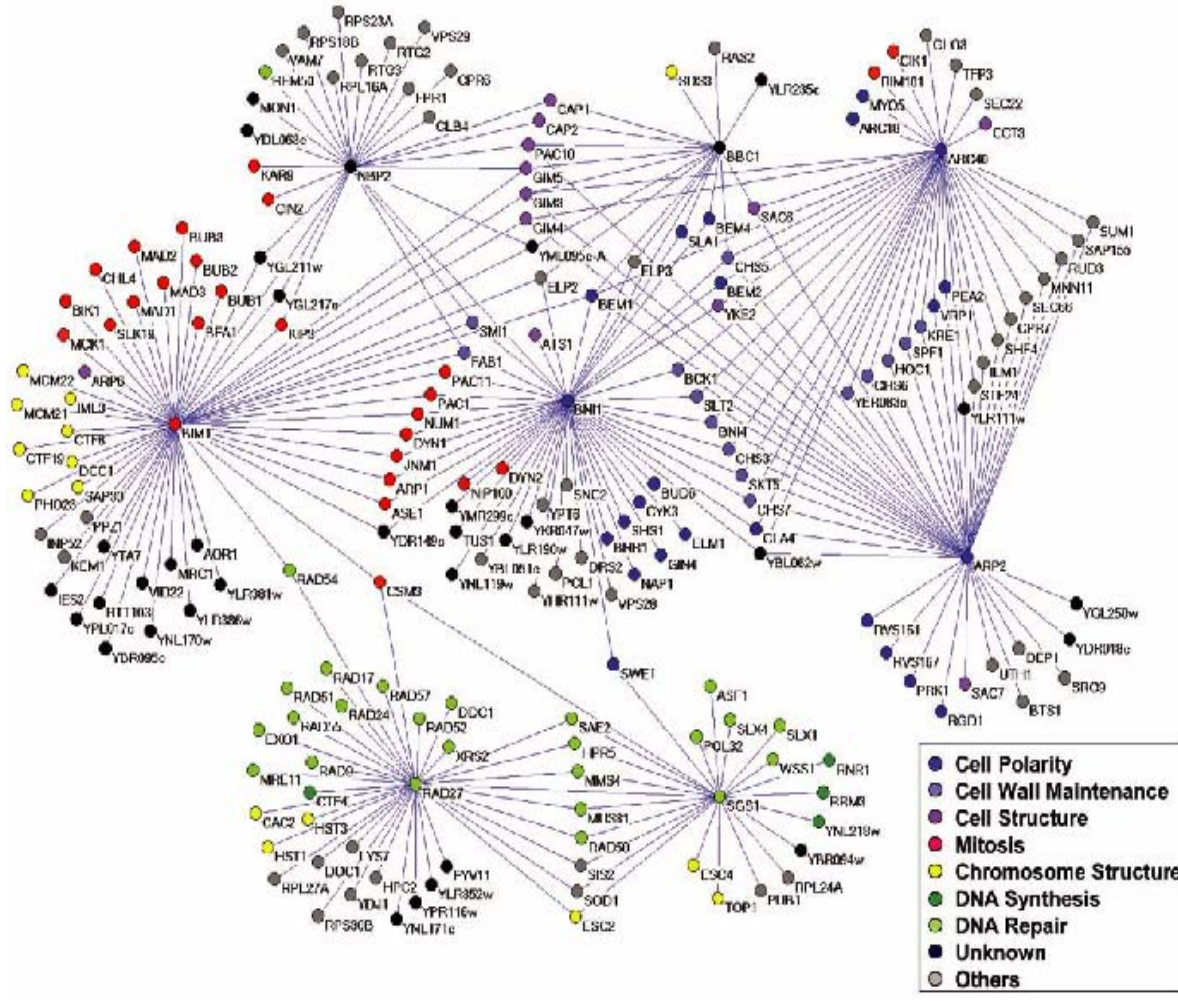
Suthram et al. *Nature* 2005  
La Count et al. *Nature* 2005

# Finding physical pathways to explain genetic interactions

## Genetic Interactions:

- Classical method used to map pathways in model species
- Highly analogous to multi-genic interaction in human disease and combination therapy
- Thousands are being uncovered through systematic studies

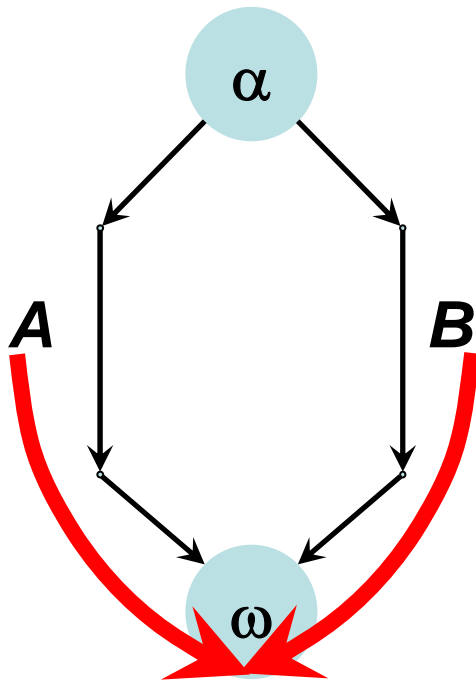
Thus as with other types, the number of known genetic interactions is *exponentially increasing...*



Adapted from Tong *et al.*, *Science* 2001

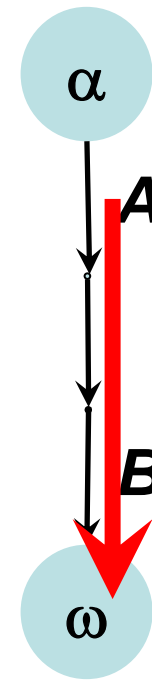
# Interpretation of genetic interactions (Guarente *T.I.G.* 1990)

## Parallel Effects (Redundant or Additive)



Single A or B mutations typically abolish their biochemical activities

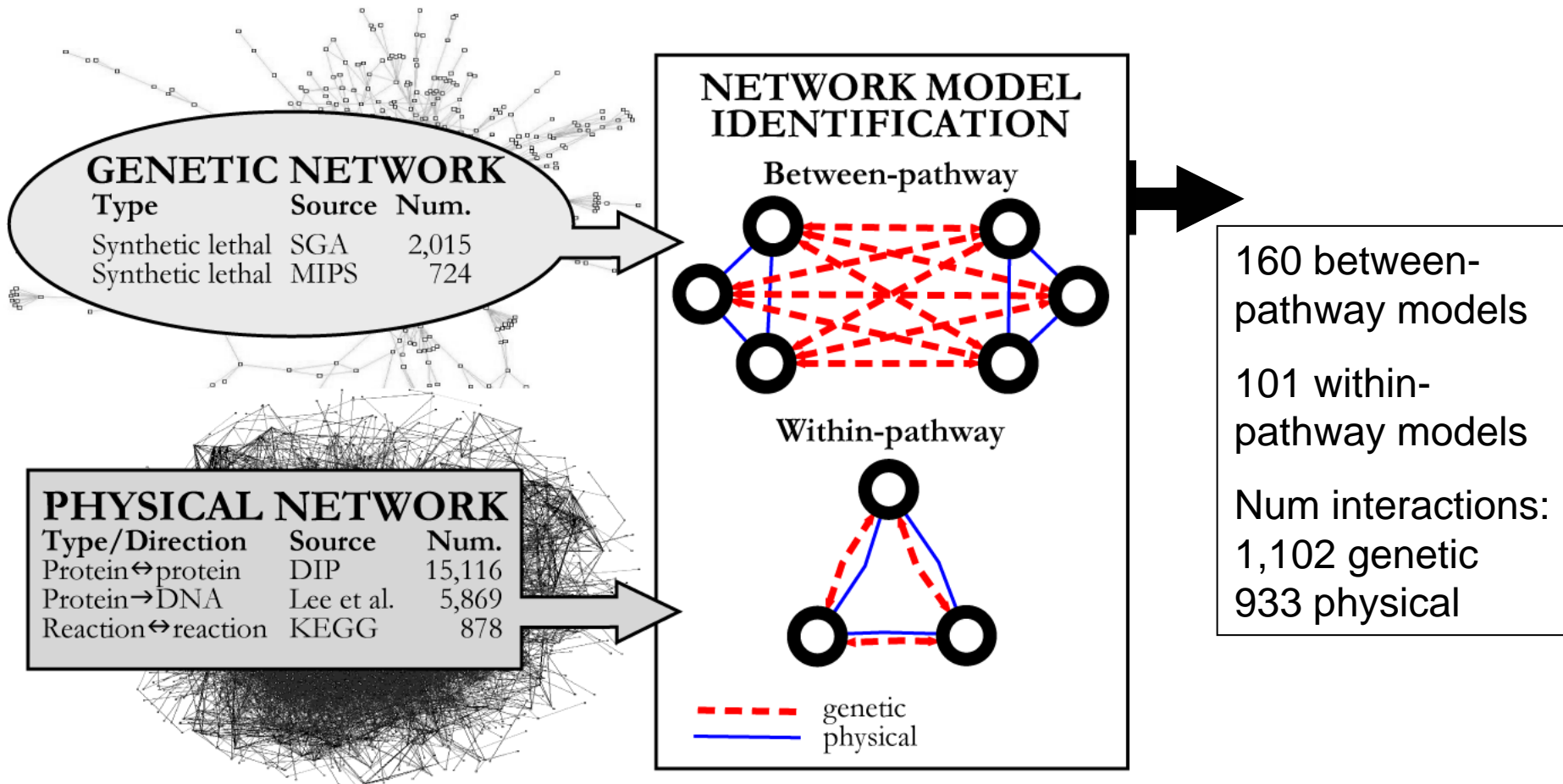
## Sequential Effects (Additive)



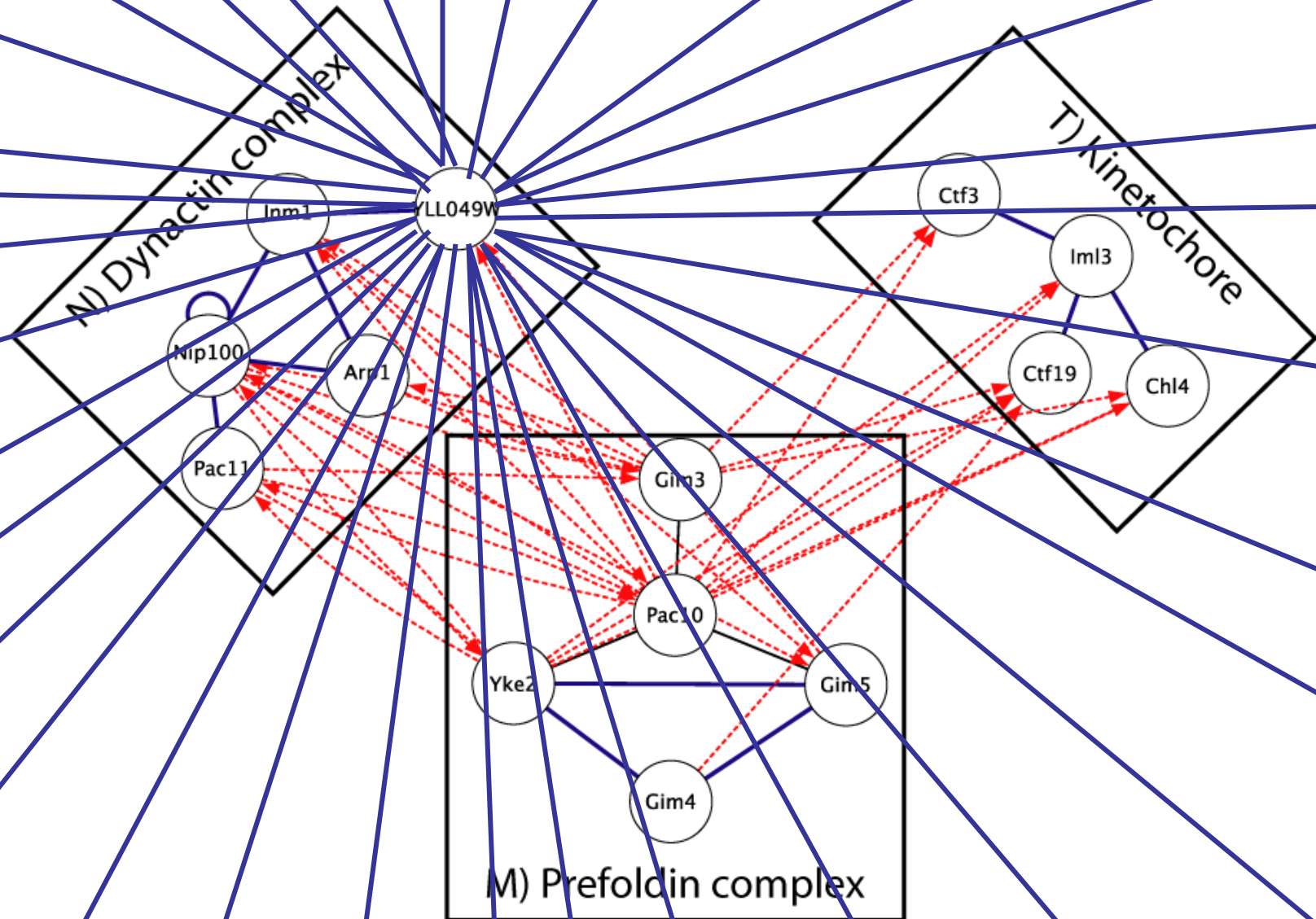
Single A or B mutations typically reduce their biochemical activities

**GOAL: Identify downstream physical pathways**

# Integration of genetic and physical interactions

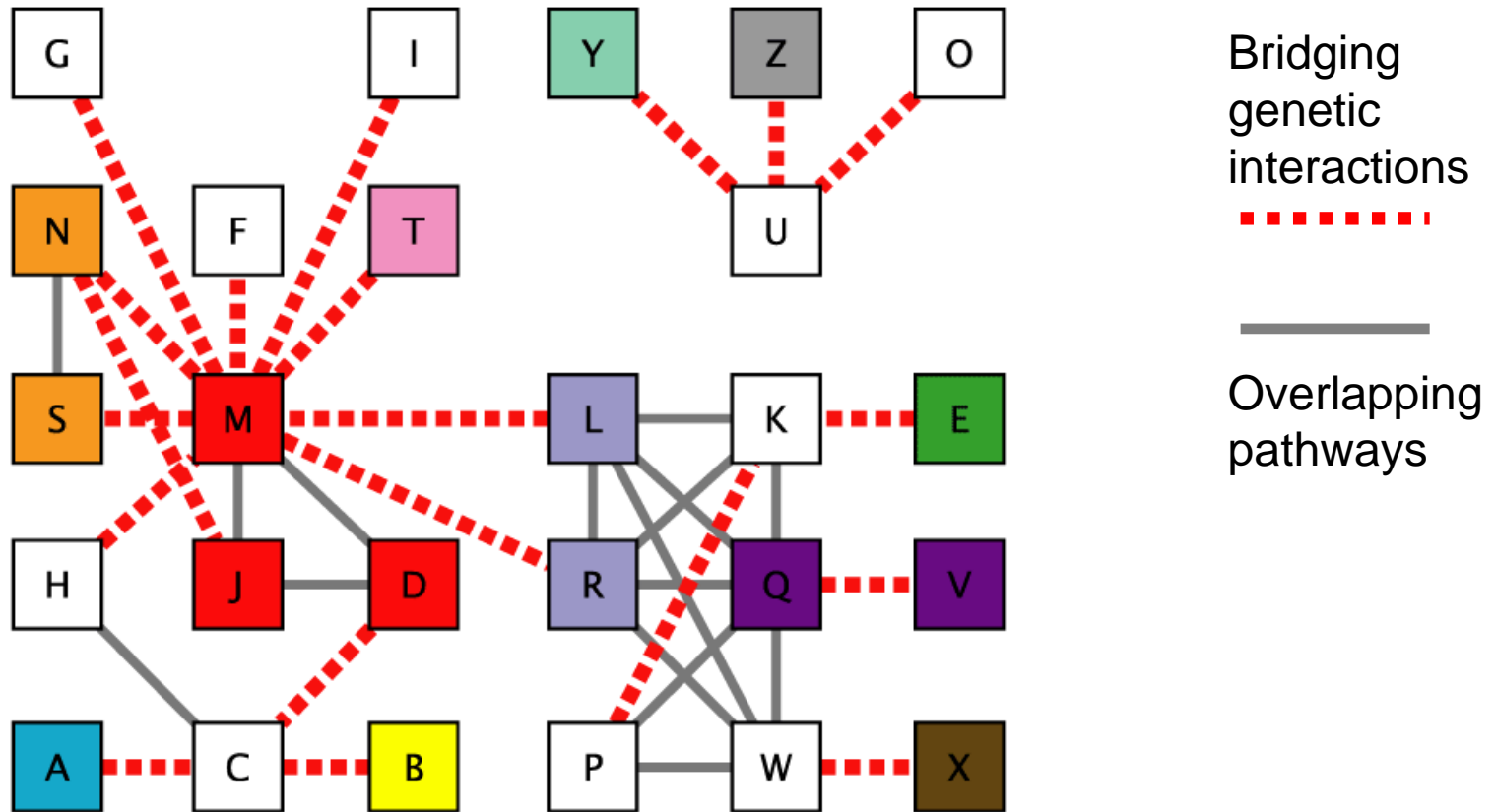


# Systematic identification of “parallel pathway” relationships in yeast





# Global organization of genetic linkages between physical pathways (A-Z)



- |                |                         |                                  |                         |                   |
|----------------|-------------------------|----------------------------------|-------------------------|-------------------|
| DNA catabolism | amino-terminal blocking | dynactin complex                 | glycoprotein metabolism | prefoldin complex |
| budding        | cell cortex             | regulation of biological process | retromer complex        | chromosome        |
| motor activity |                         |                                  |                         |                   |

OPEN SOURCE Java-based platform for modeling large molecular interaction networks

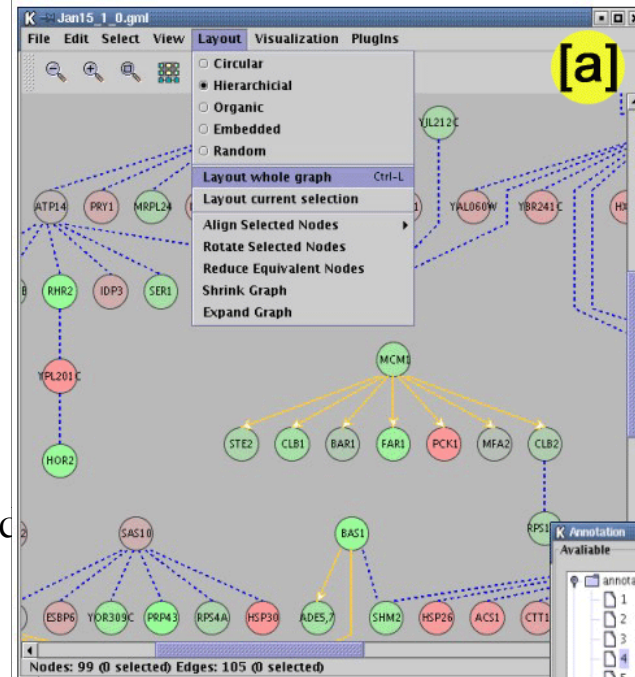
## CORE (layout & data integration)

- Layout of large interaction networks (including protein-protein, protein-DNA, genetic, and biochemical)
- Links to functional attributes (user defined GO, KEGG) and lower-level modeling environments (SBML/SBW)

- Generalized attribute-to-visual mapping

## PLUG-INS (computation, extensions)

- Pathway/network analyses implemented through an extensible PlugIn architecture



[b]

[c]

expression	myView2	myView	species
canonicalName	Name	GO Biological Process (level 6)	expression
YIL157C	FAR1	protein kinase inhibitor	-0.803
YCR108W	CLB1	cyclin-dependent protein kinase, r...	-0.566
YPR119W	CLB2	cyclin-dependent protein kinase, r...	-0.279
YMR043W	MCM1	RNA polymerase II transcription fac...	-0.654
YFLO26W	STE2	G-protein coupled receptor	-0.396
YILO15W	BAR1	aspartic-type endopeptidase	-0.207
YNL145W	MFA2		-0.098
YKR097W	PCK1	carboxy-lyase   phosphoenolpyruv...	1.224
YDR025W	RPS11A	structural constituent of ribosome	-0.39
YLR255W	GSY2	transferase, transferring hexosyl gr...	0.4
YGL255W	GSY1	transferase, transferring hexosyl gr...	0.676
YGL255W	GLG2	transferase, transferrin hexosyl tr...	0.329

[d]

JOINT PROJECT with the Inst. for Systems Biology (Hood), Inst. Pasteur (Schwikowski), Sloan-Kettering (Sander), U. Toronto (Bader), UCSF (Conklin) and Agilent (Adler/Kuchinsky)

# Transcriptional response of *Saccharomyces cerevisiae* to DNA-damaging agents does not identify the genes that protect against these agents

Geoff W. Birrell\*, James A. Brown\*, H. Irene Wu\*, Guri Giaever†, Angela M. Chu†, Ronald W. Davis†, and J. Martin Brown\*\*

Departments of \*Radiation Oncology and †Biochemistry, Stanford University School of Medicine, Stanford, CA 94305

Contributed by Ronald W. Davis, May 8, 2002

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type parental strain to the same DNA-damaging agents. We found no relationship between the genes necessary for survival to the DNA-damaging agents and those genes whose transcription is increased after exposure. These data show that few, if any, of the genes involved in repairing the DNA lesions produced in this study, including double-strand breaks, pyrimidine dimers, single-strand breaks, base damage, and DNA cross-links, are induced in response to toxic doses of the agents that produce these lesions. This finding suggests that the enzymes necessary for the repair of these lesions are at sufficient levels within the cell. The data also suggest that the nature of the lesions produced by DNA-damaging agents cannot easily be deduced from gene expression profiling.

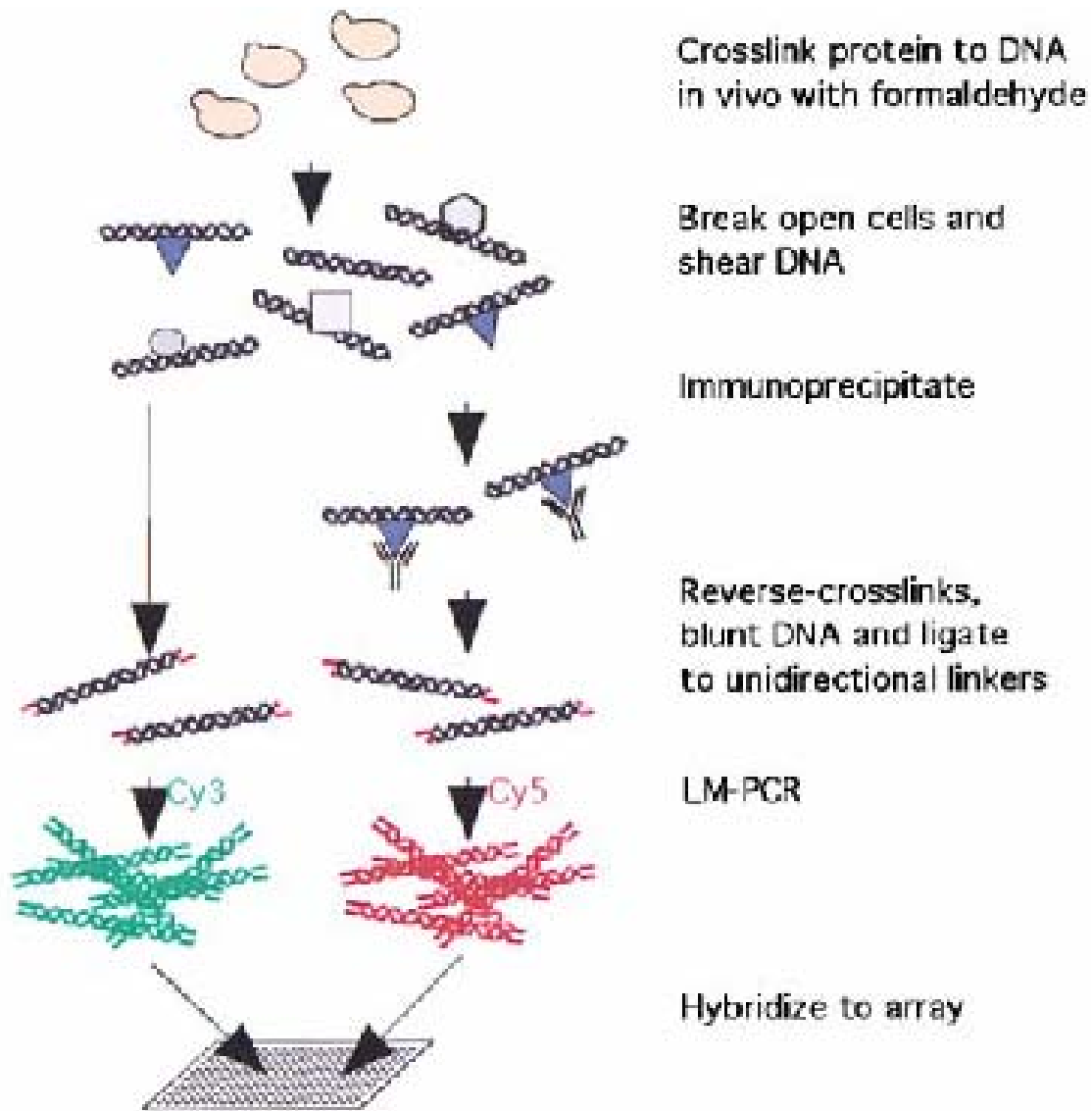
Can this apparent paradox be explained by a physical model of the DNA damage response?

this hypothesis.

Deletion of the genes has been accomplished by an international consortium, the *Saccharomyces* Genome Deletion Project, that has replaced all of the ≈6,200 known open reading frames (ORFs) of yeast by using a PCR-mediated gene deletion strategy (20). In addition to a selectable marker, two molecular bar codes or “tags,” unique 20-base oligonucleotide sequences, are in the replacement cassette. These tags, after PCR amplification, can be detected by hybridization to the corresponding complementary sequence in a high-density oligonucleotide array, thus enabling the relative abundances of each tag, and hence the abundances of each deletion strain, to be determined (20). We have recently shown that this system can detect essentially all of

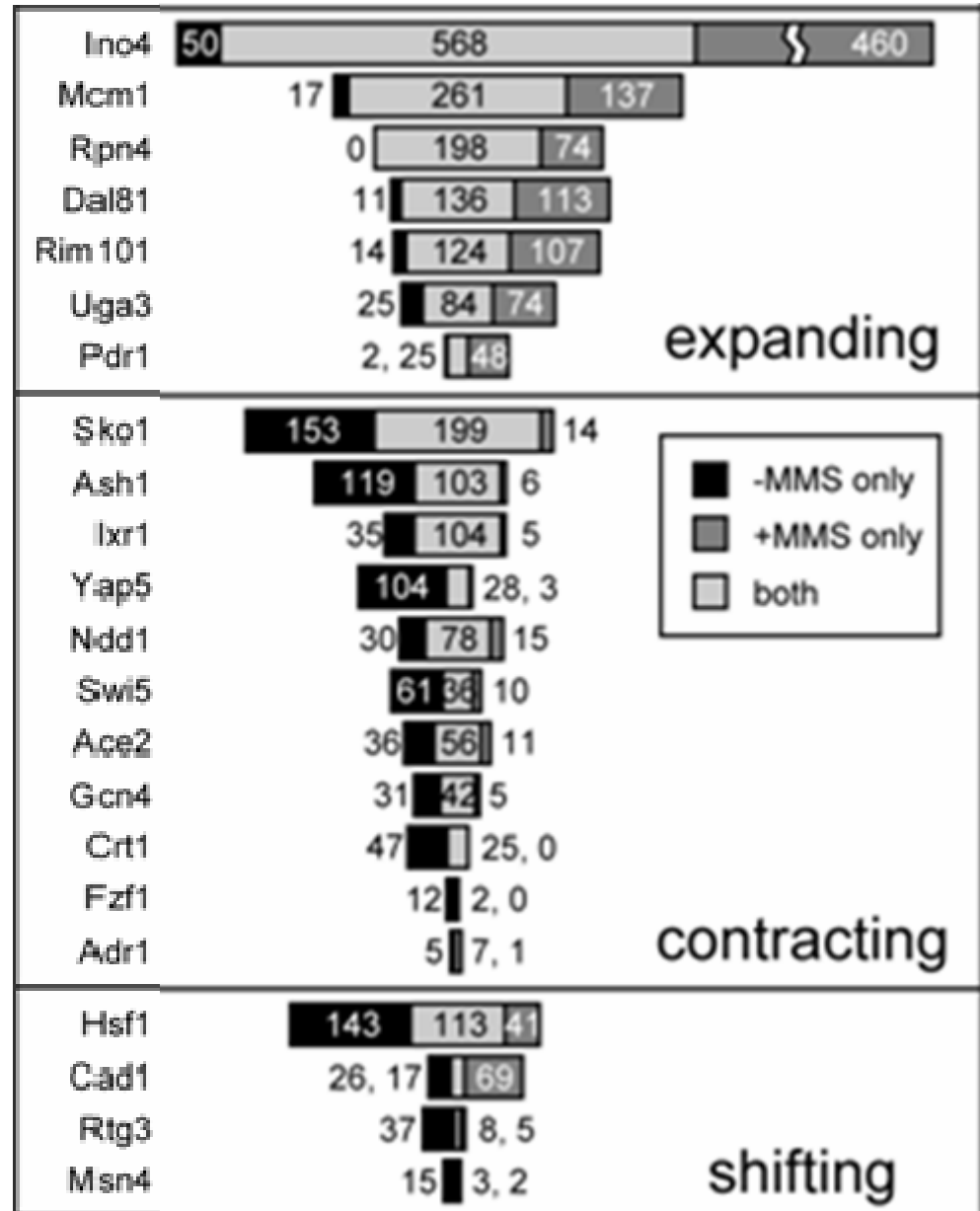
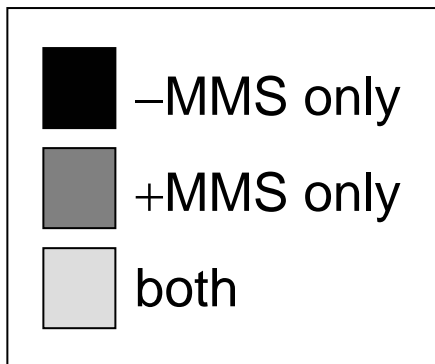
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# ChIP-chip measurement of protein→DNA interactions

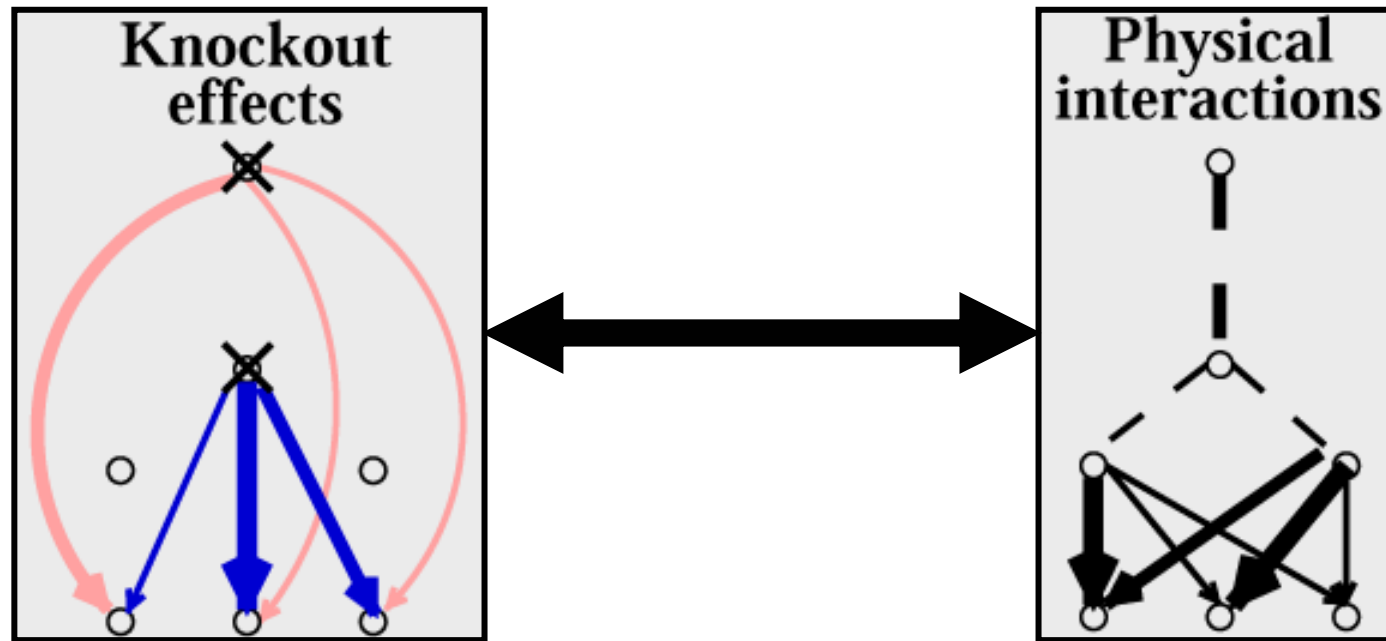




# A systems approach to mapping DNA damage networks

Numbers of promoters bound by each of 30 transcription factors (TFs) before and after exposure to methyl-methane sulfonate (MMS)



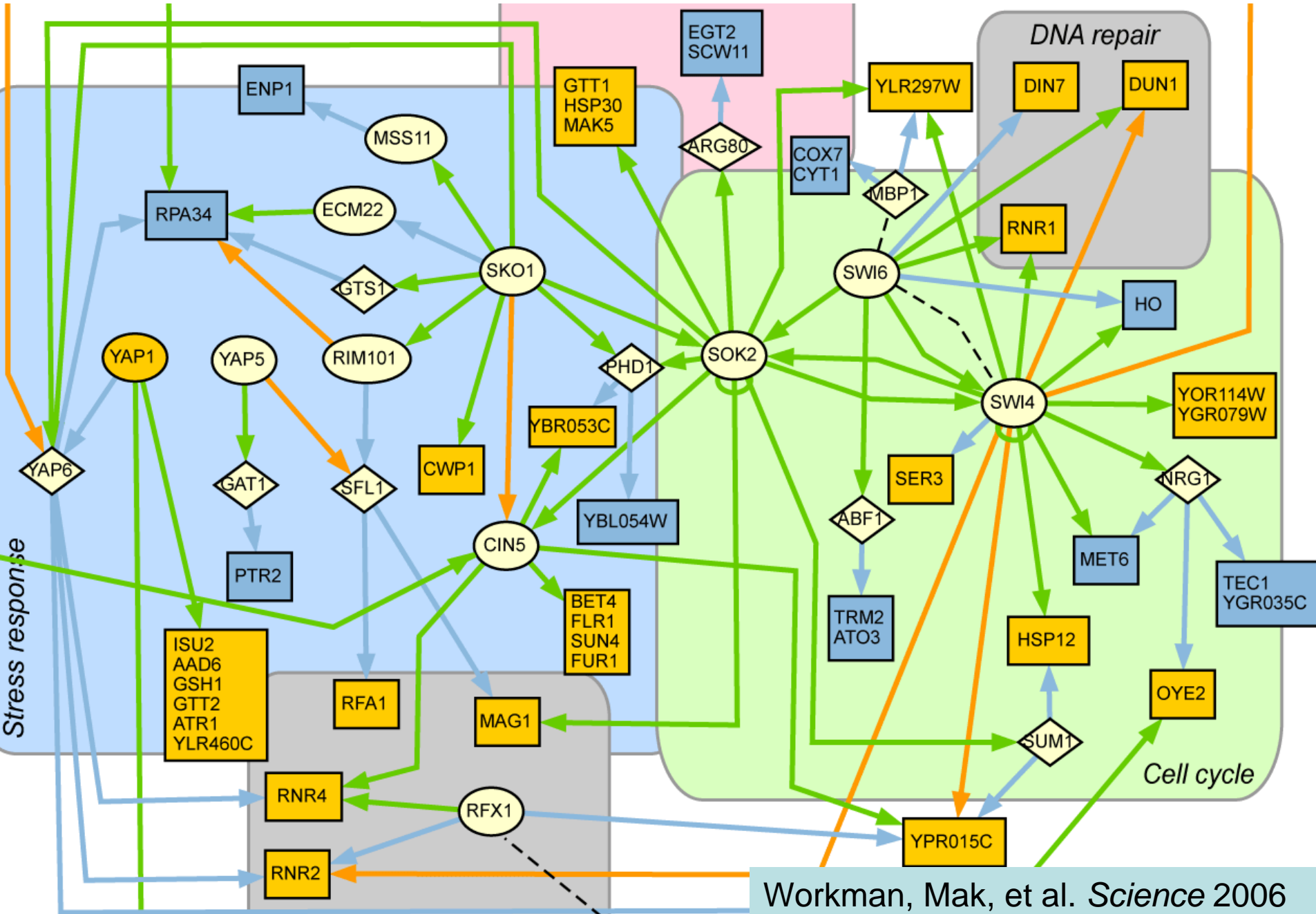
# Validation of physical network by systematic gene knockout analysis



 Knockout causes up-regulation  
 Knockout causes down-regulation

 TF-promoter binding  
 Protein-protein binding

*Validation of binding with knockout data yields a large regulatory network*



# Transcriptional response of *Saccharomyces cerevisiae* to DNA-damaging agents does not identify the genes that protect against these agents

Geoff W. Birrell\*, James A. Brown\*, H. Irene Wu\*, Guri Giaever†, Angela M. Chu†, Ronald W. Davis†, and J. Martin Brown\*\*

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Contributed by Ronald W. Davis, May 8, 2002

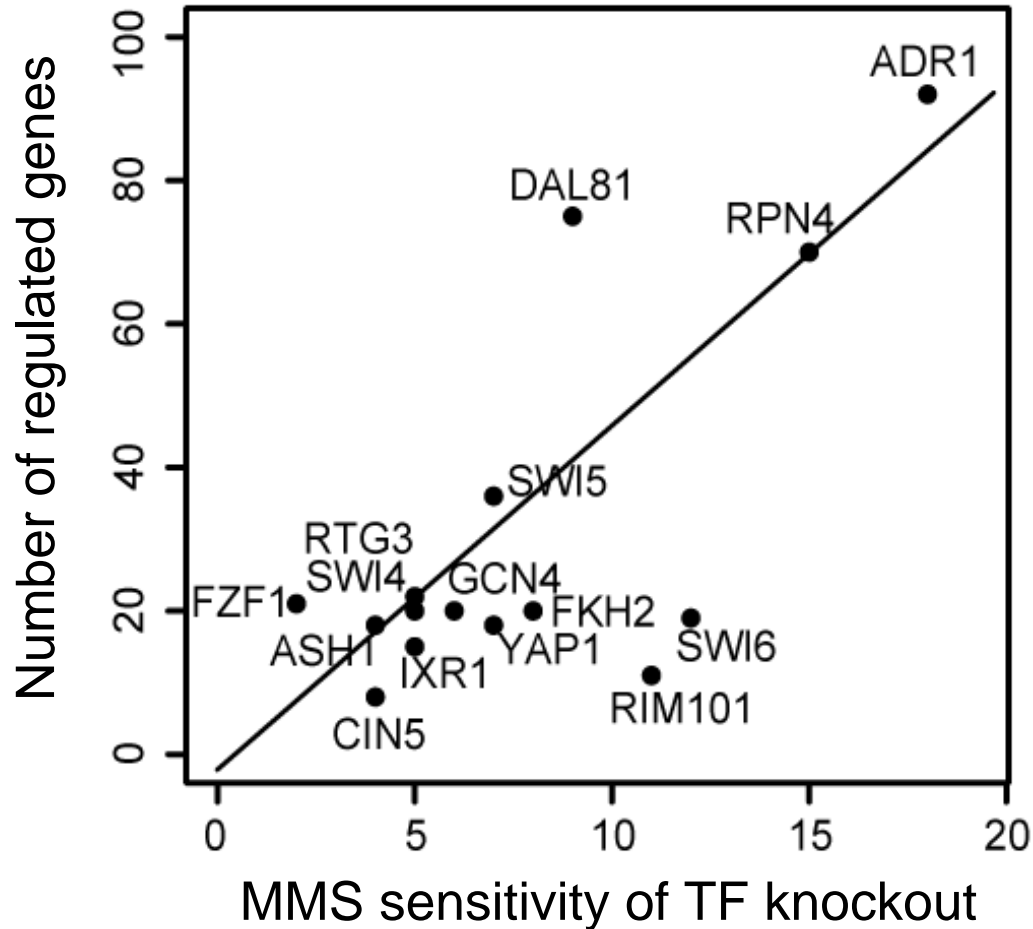
The recent completion of the deletion of all of the nonessential genes in budding yeast has provided a powerful new way of determining those genes that affect the sensitivity of this organism to cytotoxic agents. We have used this system to test the hypothesis that genes whose transcription is increased after DNA damage are important for the survival to that damage. We used a pool of 4,627 diploid strains each with homozygous deletion of a nonessential gene to identify those genes that are important for the survival of yeast to four DNA-damaging agents: ionizing radiation, UV radiation, and exposure to cisplatin or to hydrogen peroxide. In addition we measured the transcriptional response of the wild-type parental strain to the same DNA-damaging agents. We found no relationship between the genes necessary for survival to the DNA-damaging agents and those genes whose transcription is increased after exposure. These data show that few, if any, of the genes involved in repairing the DNA lesions produced in this study, including double-strand breaks, pyrimidine dimers, single-strand breaks, base damage, and DNA cross-links, are induced in response to toxic doses of the agents that produce these lesions. This finding suggests that the enzymes necessary for the repair of these lesions are at sufficient levels within the cell. The data also suggest that the nature of the lesions produced by DNA-damaging agents cannot easily be deduced from gene expression profiling.

conferring resistance to that agent, and hence provide information on its mechanism. Recent publications have, in fact, suggested that several of the genes induced by DNA-damaging agents are involved in the repair of DNA damage and hence in the protection of the cell against such treatments (17–19). However, the assumption that genes whose expression increases in response to a particular cytotoxic agent are those that protect against the damage caused by the agent has not been formally tested. Here we use a pool of strains of budding yeast, *S. cerevisiae*, with deletion of all nonessential genes to directly test this hypothesis.

Deletion of the genes has been accomplished by an international consortium, the *Saccharomyces* Genome Deletion Project, that has replaced all of the ≈6,200 known open reading frames (ORFs) of yeast by using a PCR-mediated gene deletion strategy (20). In addition to a selectable marker, two molecular bar codes or “tags,” unique 20-base oligonucleotide sequences, are in the replacement cassette. These tags, after PCR amplification, can be detected by hybridization to the corresponding complementary sequence in a high-density oligonucleotide array, thus enabling the relative abundances of each tag, and hence the abundances of each deletion strain, to be determined (20). We have recently shown that this system can detect essentially all of

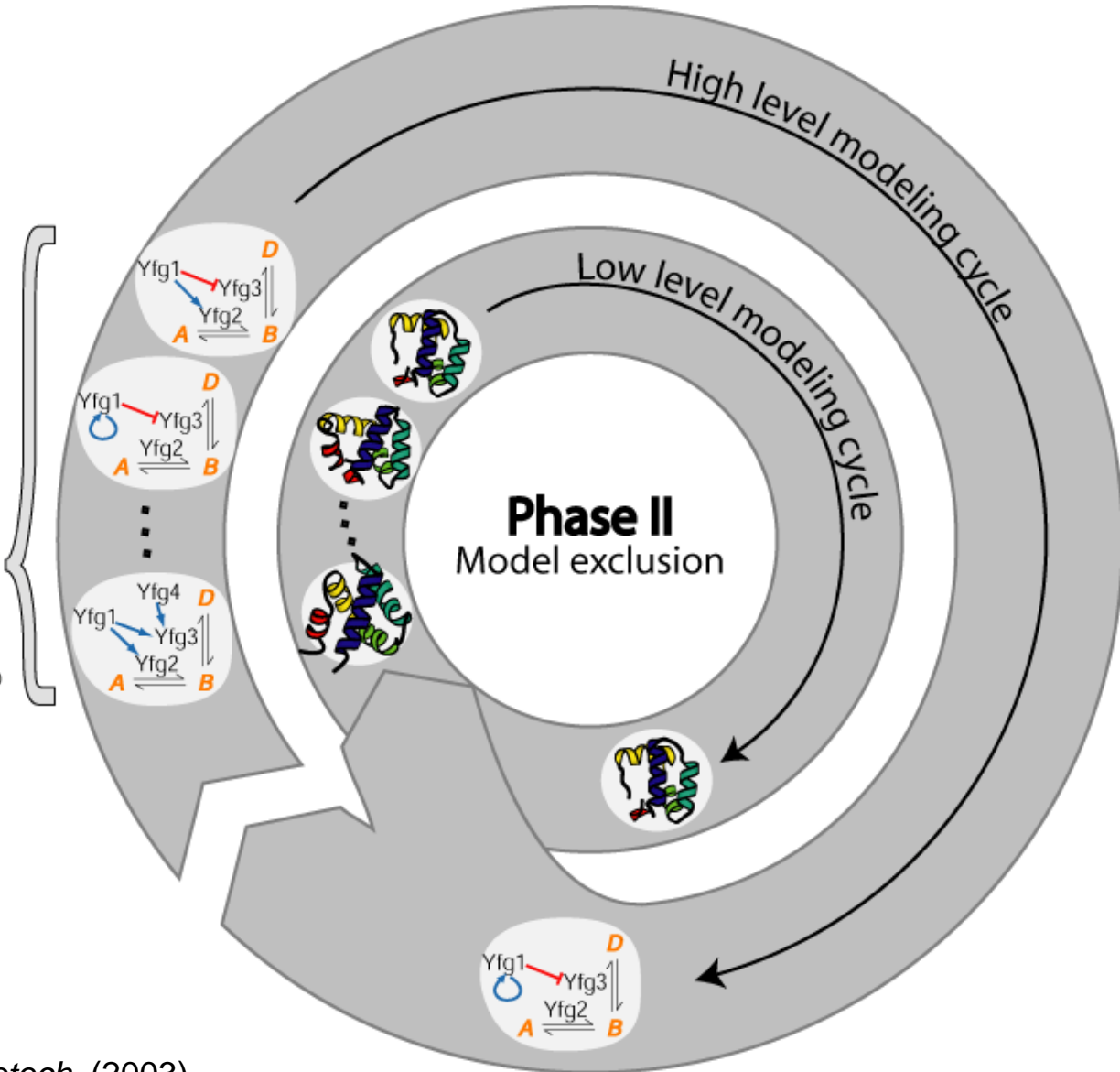
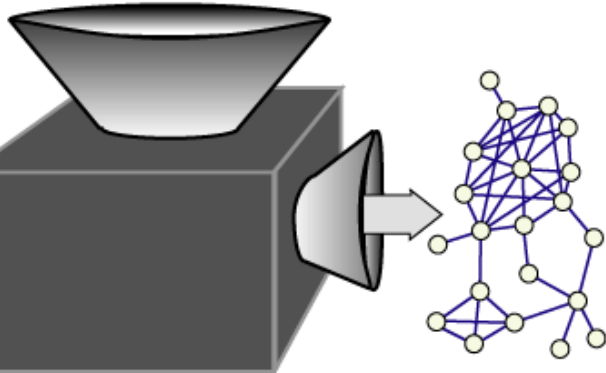


Sensitivity of the TF knockout phenotype correlates with its number of regulated targets



# The vision: First build the scaffold, then add the details

## Phase I Model generation



## Network Comparison:

Sourav Bandyopadhyay,  
Ryan Kelley,  
Silpa Suthram



**Collaborators:** Roded Sharan (Tel Aviv), Richard Karp (Berkeley)

## DNA Damage Networks:

Craig Mak  
Chris Workman



## Collaborators:

Leona Samson (MIT)  
Tom Begley (U Albany)

**Funding:** Packard Fellowship; NIEHS, NCRR, NIGMS, NIAID, NSF, Unilever

**Websites:** [www.pathblast.org](http://www.pathblast.org); [www.cytoscape.org](http://www.cytoscape.org)