

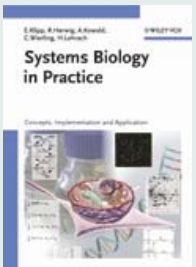
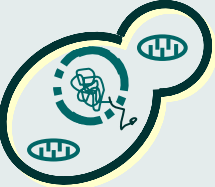
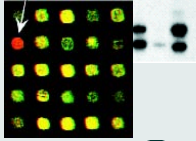
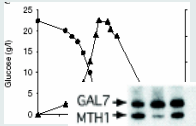
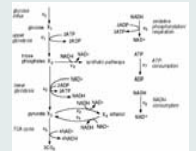
ICSB 2006, Yokohama, Oct 9-13, 2006

Dynamic Modeling of Stress Response of Yeast Cells - Timing of Events

Edda Klipp
Max Planck Institute for Molecular Genetics
Innestr. 73, 14195 Berlin, Germany
http://www.molgen.mpg.de/~ag_klipp



$$\begin{aligned} dX_1/dt &= v_0 - v_1 \\ dX_2/dt &= 2v_1 - v_2 - v_9 \\ dX_3/dt &= v_2 - v_3 + v_4 - v_5 \\ dX_4/dt &= v_3 - v_4 \\ dNADH/dt &= v_2 - v_5 + v_4 + 4v_5 - v_6 \\ dATP/dt &= -2v_1 + 2v_2 + 3v_6 - v_7 \end{aligned}$$



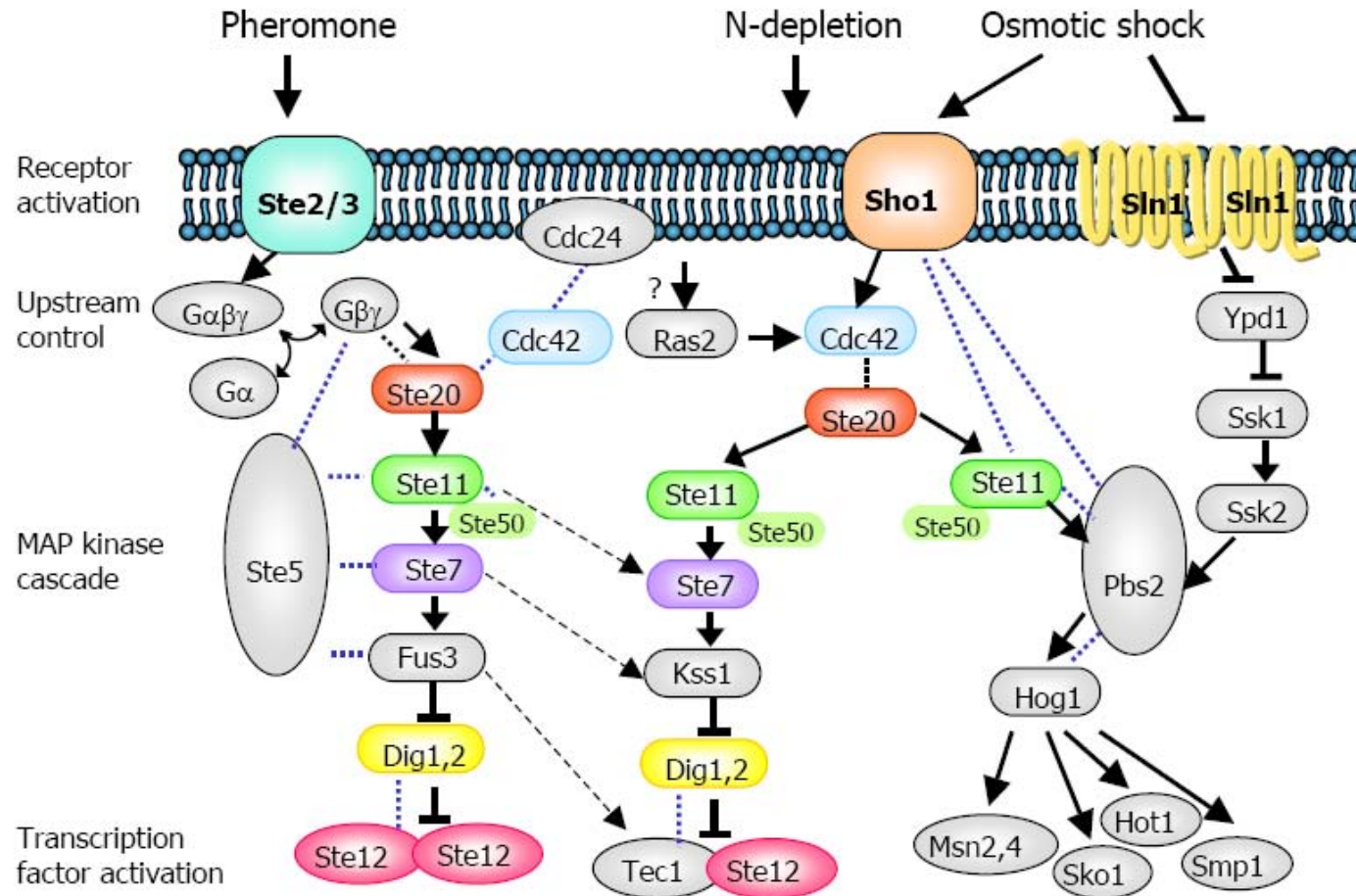


Timing of Stress Response

- **Response to Osmotic Stress**
- **Crosstalk of Pheromone Pathway and Filamentous Growth Pathway**
- **Cell Cycle and its Regulation by Signaling Pathways**



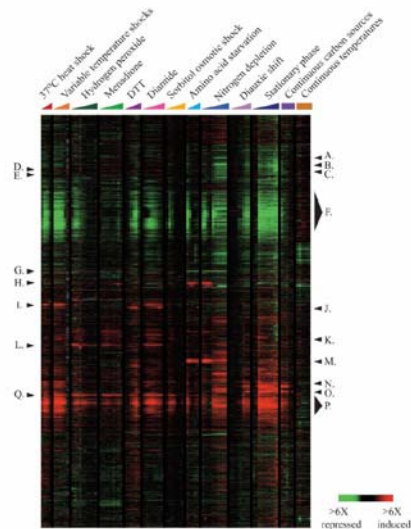
Signaling Pathways in Baker's Yeast





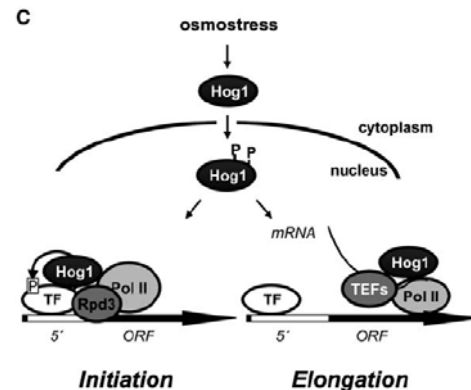
Experiments w.r.t. Stress Response

Stress response on transcriptional level



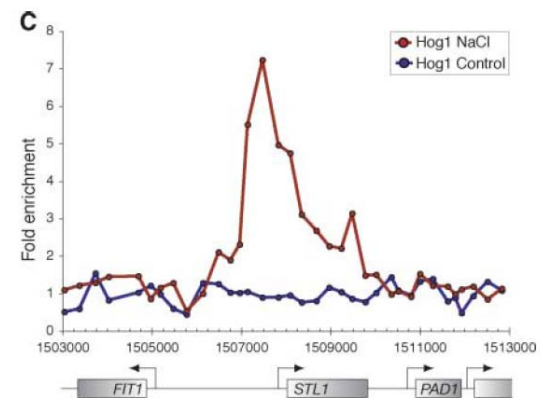
Gasch et al., 2000, Mol Cell Biol

Stress-activated Hog1 – a selective transcription elongation factor for genes responding to osmotic stress



Proft et al., 2006, Molecular Cell

Activated Signal Transduction Kinases Frequently Occupy Target Genes

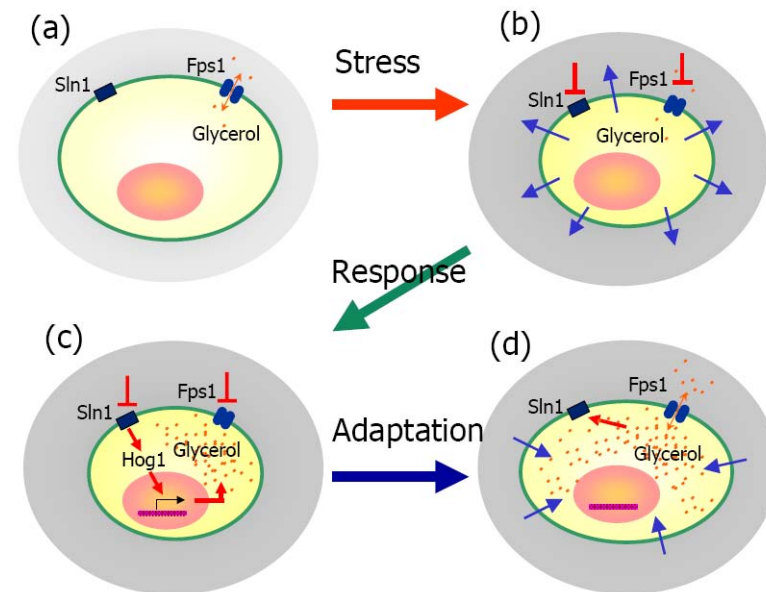


Pokholok, ..., Young, 2006, Science



Yeast: Response to Osmotic Stress

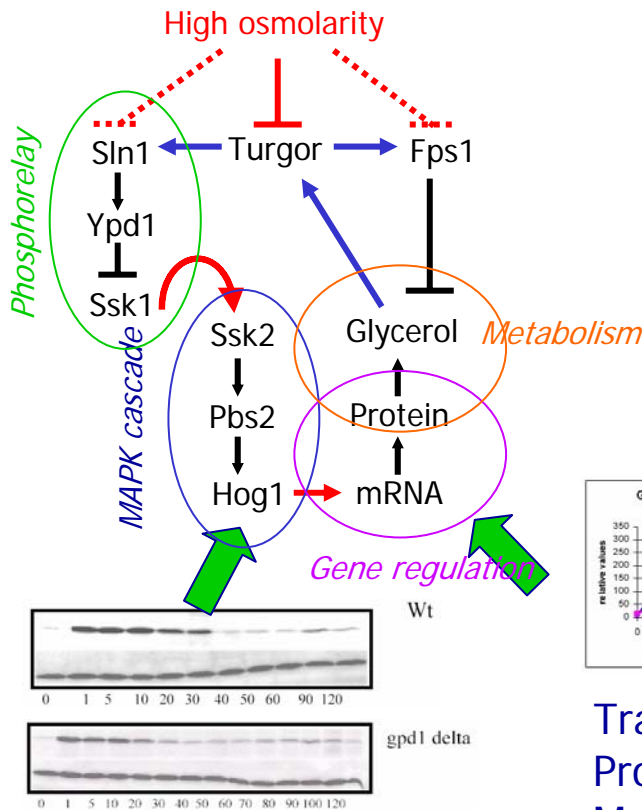
- Active regulation of cellular volume
- Accumulation of glycerol as osmolyte
- Closure of the aquaglyceroporin Fps1 (a glycerol channel)
- „Genome remodeling“ – global change of expression pattern
- Regulation of cell cycle progression





Modeling Pipeline

Construct network from literature data and experts' knowledge

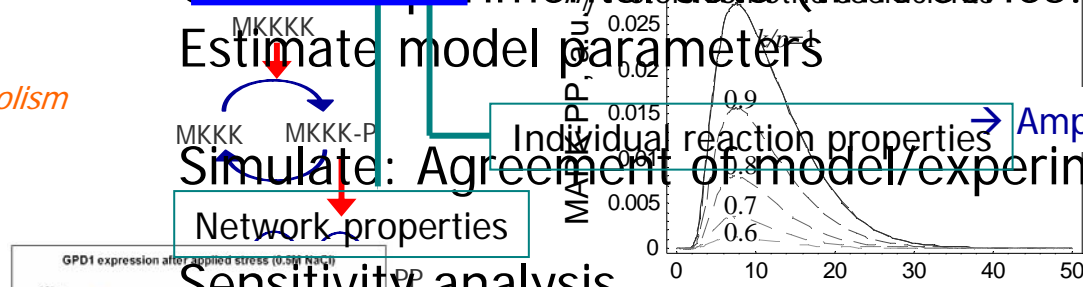


Study properties of small modules, e.g. MAPK cascade, G-protein cycles, ...

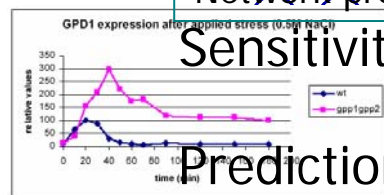
$$\frac{dS_i}{dt} = \sum_{j=1}^n n_{ij} v_j$$

S_i – metabolite concentrations
 v_j – reaction rates
 n_{ij} – stoichiometric coefficients

Collect experimental data (time series!!!)
 Estimate model parameters
 Simulate: Agreement of model/experiment?



Network properties
 Sensitivity analysis

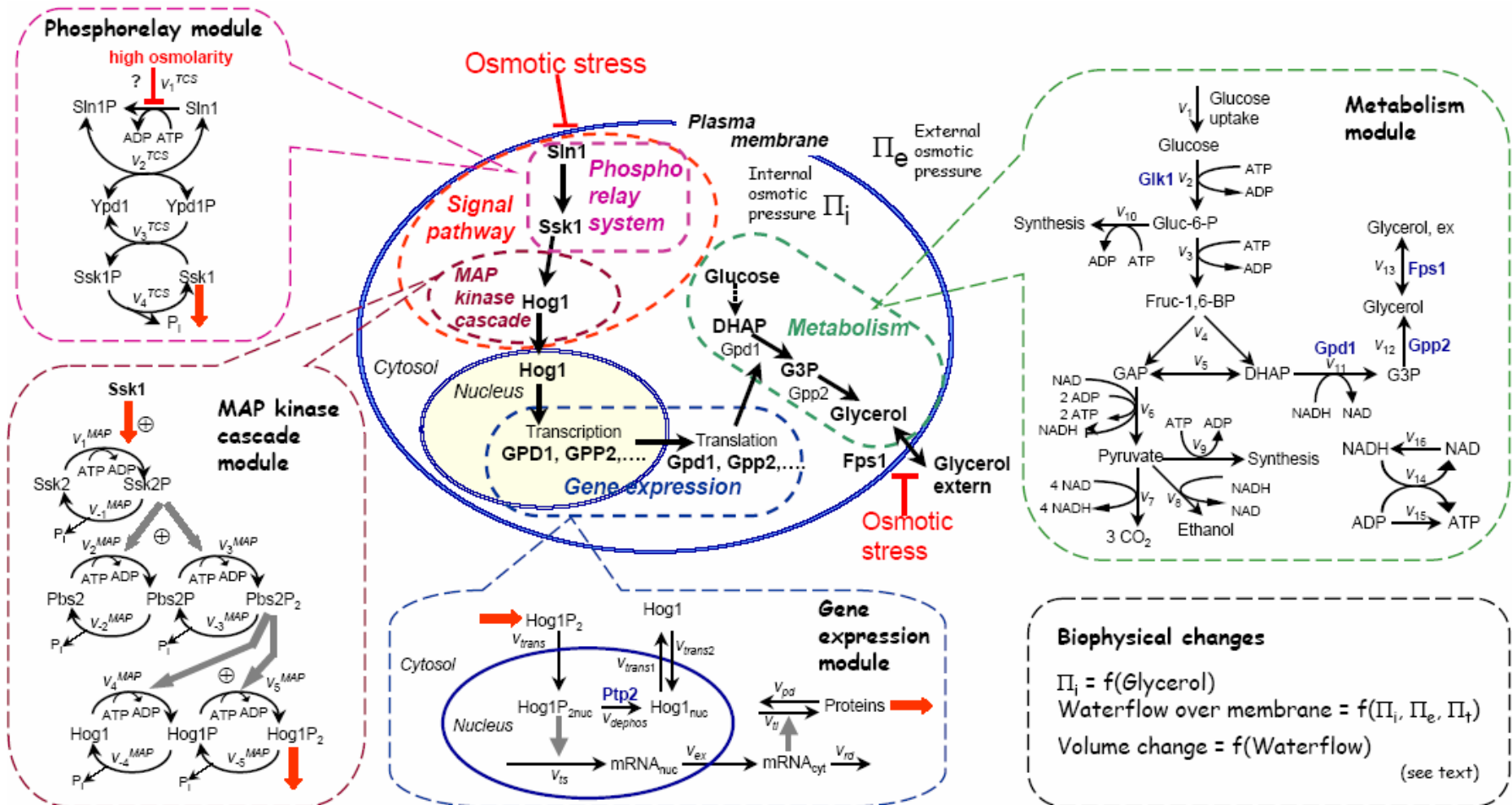


Prediction of hitherto untested scenarios

- Deletion mutants
 - Transcriptome data – mRNA levels
 - Proteome data – phosphorylation concentration changes
 - Metabolome data – metabolite concentration changes
 - New experimental scenarios



Osmostress Response – Full Model

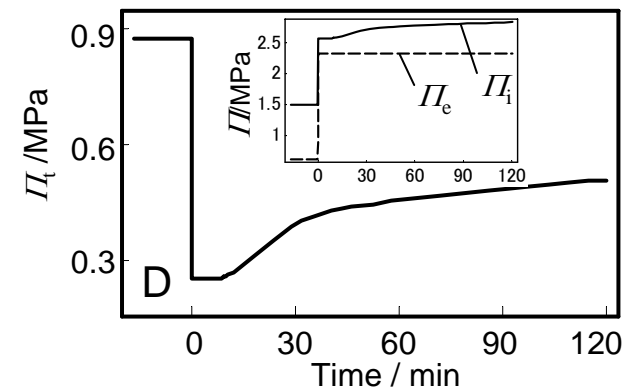
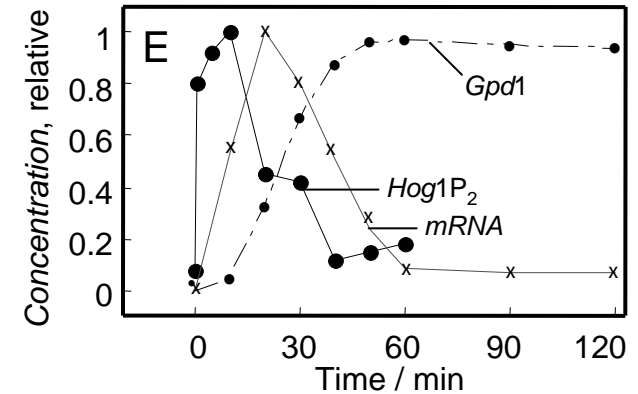
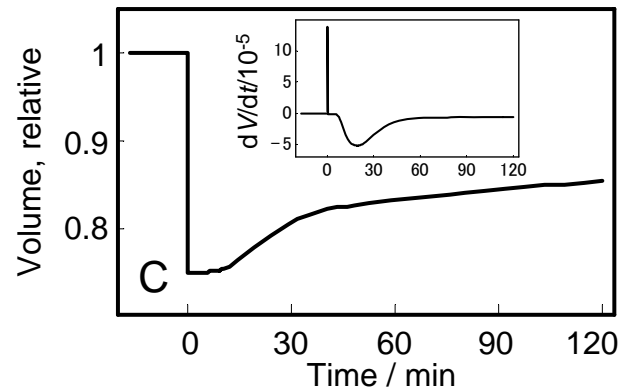
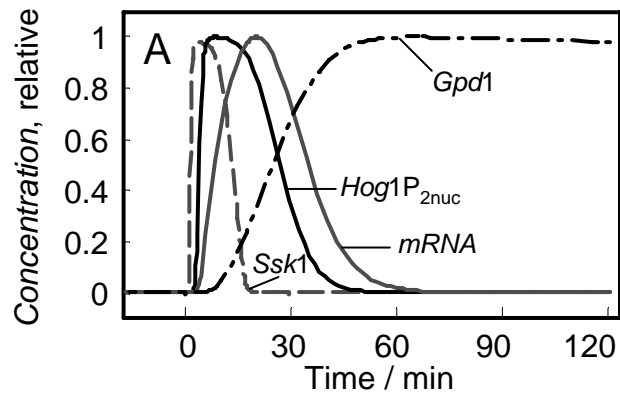
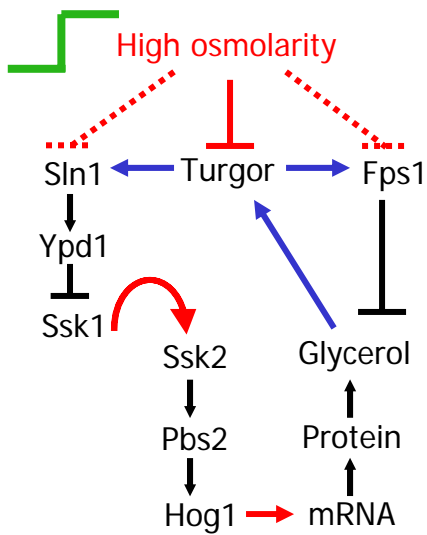


Klipp, Nordlander, Krüger, Gennemark & Hohmann, *Nature Biotechnol.*, 2005



The Standard Experiment

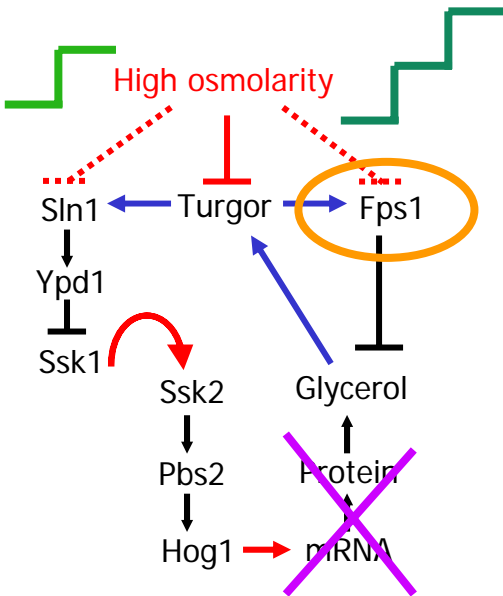
Wild Type Cells, shock with 0.5 M NaCl



Klipp et al., *Nature Biotechnol.*, 2005



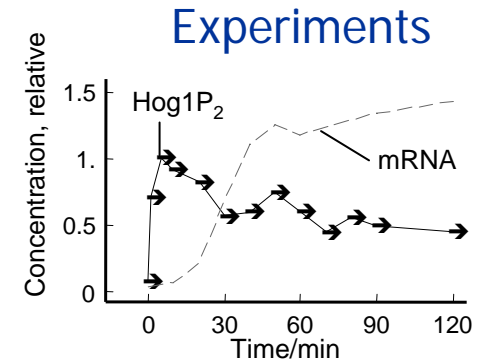
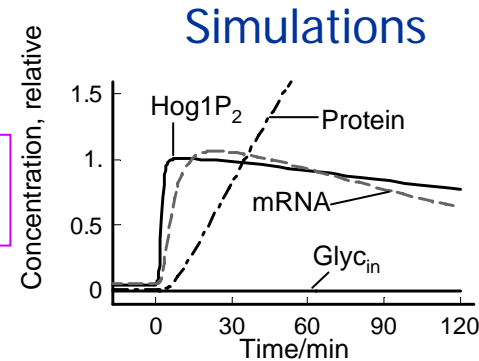
Osmotic stress model: Test cases



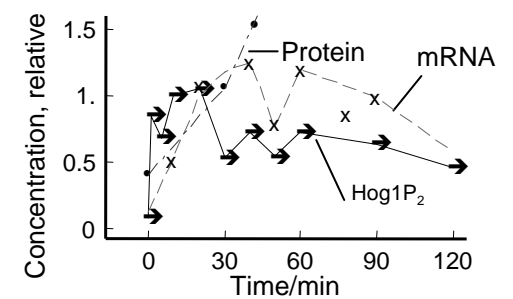
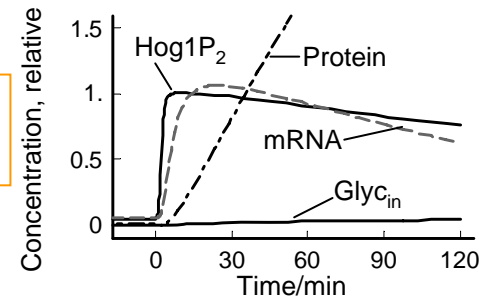
gpd1 Δ gpd2 Δ
mutant

Fps1 open,
constitutively

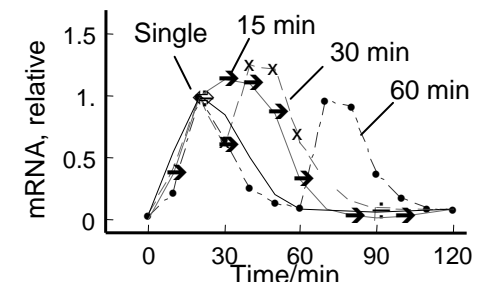
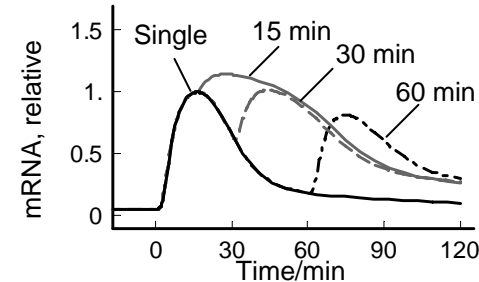
Repeated
osmstress



No enzyme production – no glycerol production – no pathway downregulation



Open Fps1 – reduced glycerol accumulation – reduced pathway downregulation

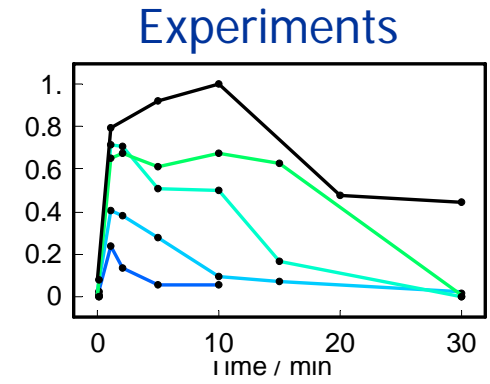
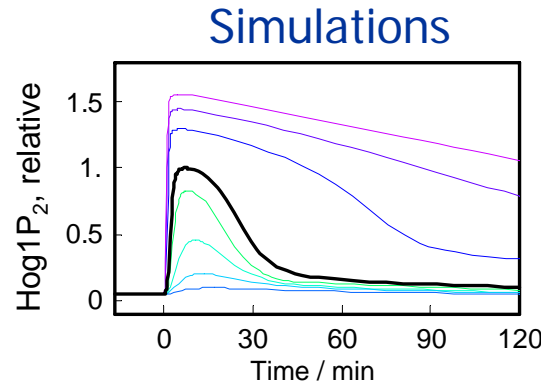
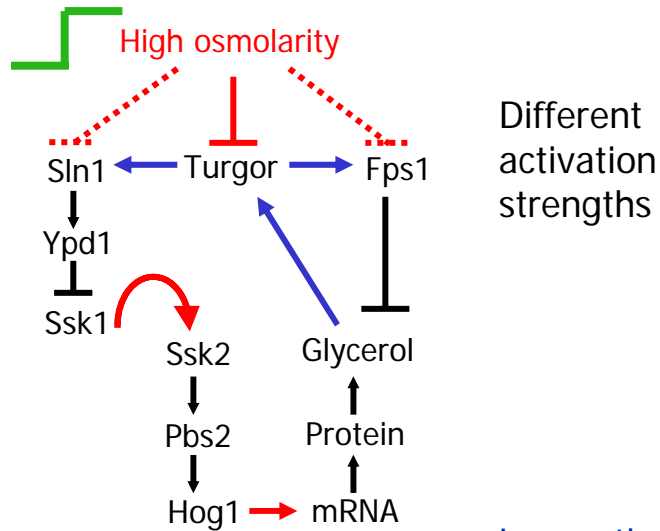


Klipp et al., *Nature Biotechnol.*, 2005

Cells are competent to respond to a second shock.



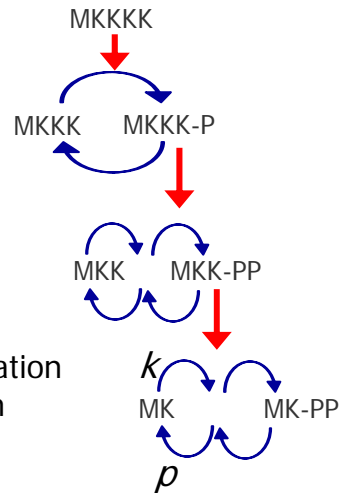
Osmotic stress: Different stress levels



NaCl / M — 0.1 — 0.3 — 0.5 — 1.0
 — 0.2 — 0.4 — 0.8 — 1.4

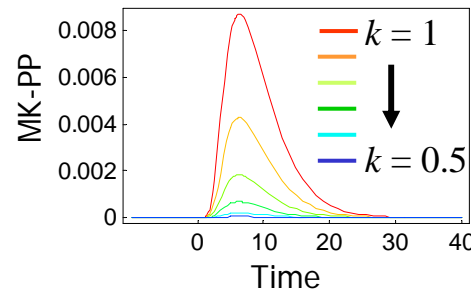
Low activation: increase of amplitude, stronger activation: increase of duration

MAP
kinase
cascade

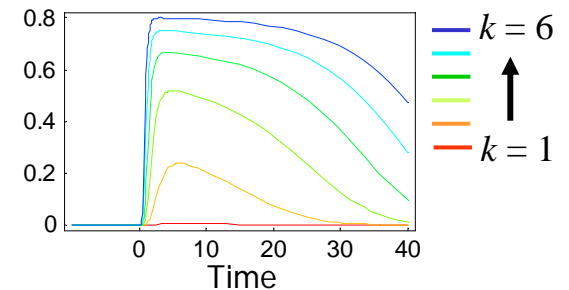


- Hierarchical regulation
- Mass conservation on each level

Low activation

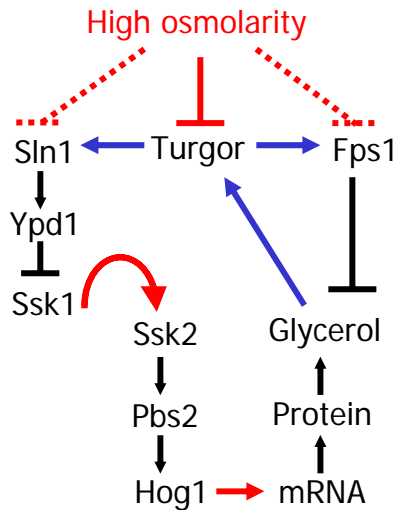


Strong activation

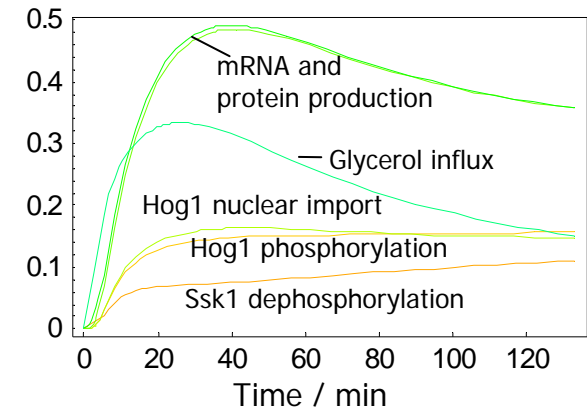
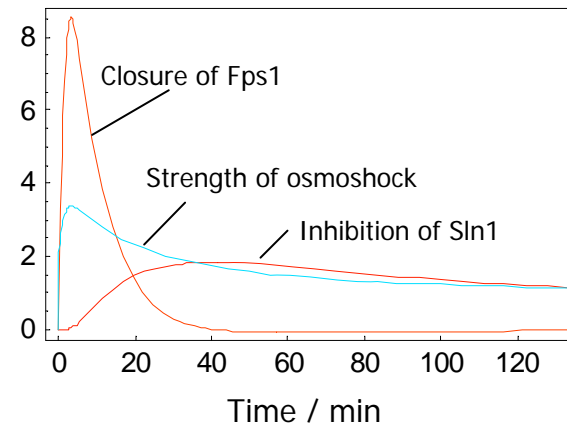




Osmotic stress response: What is the impact of specific components over time ?



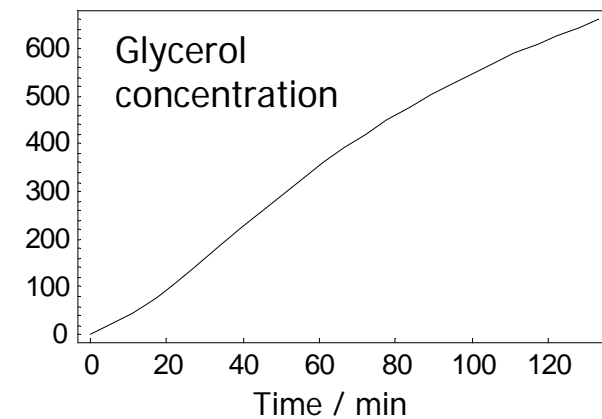
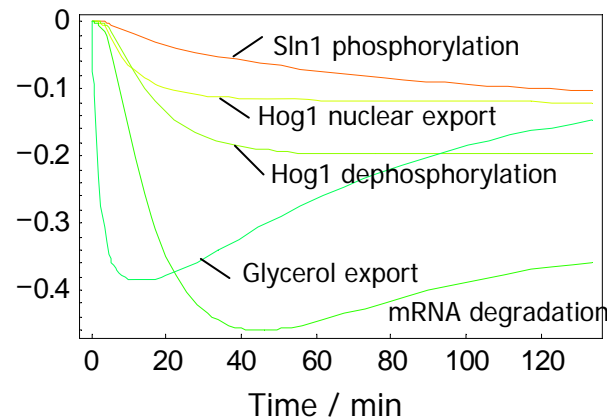
Time-dependent Response Coefficients Related to Glycerol Concentration



Response coefficients

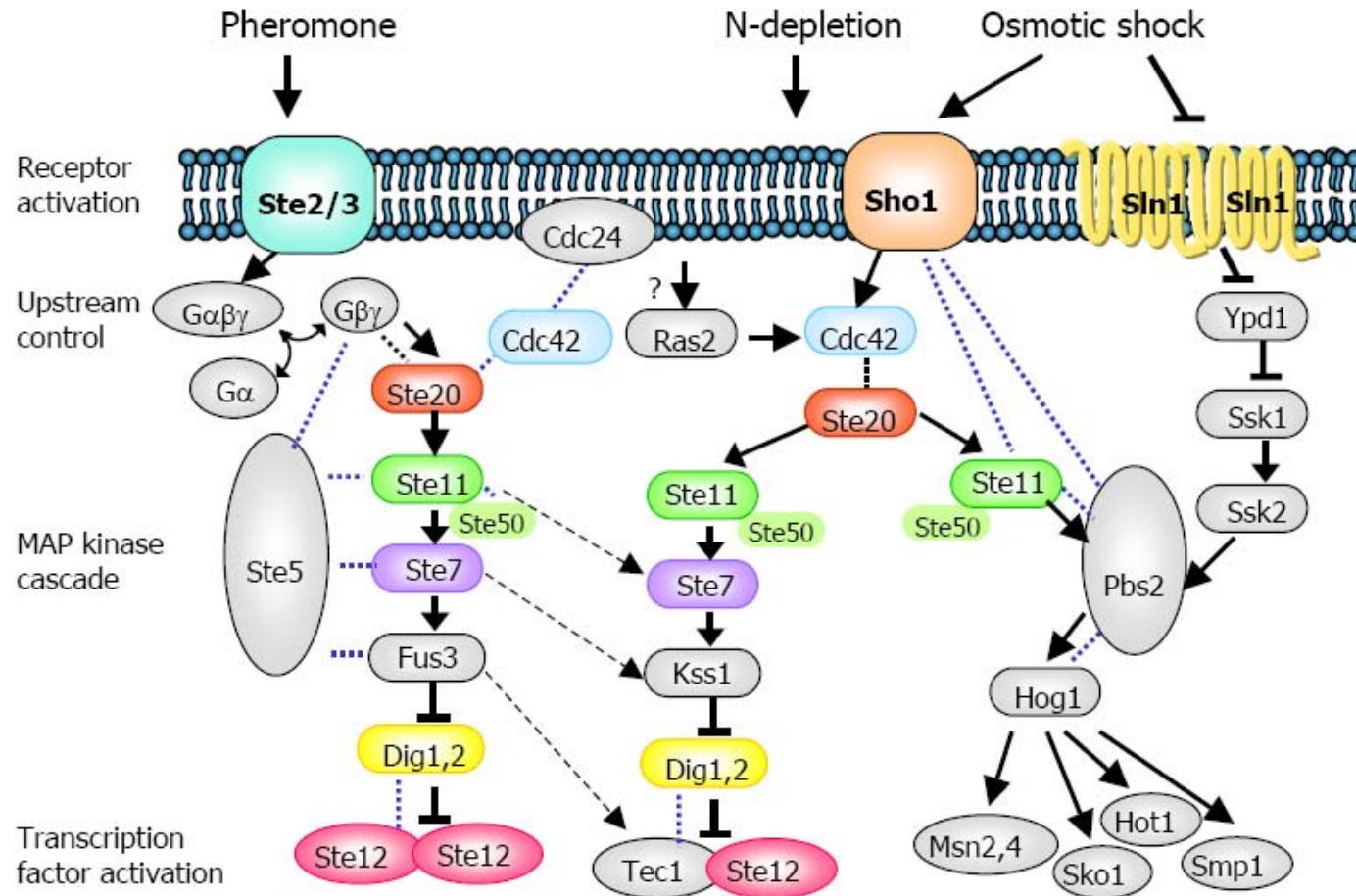
$$R_{p_l}^{S_i} = \frac{p_l}{S_i(t)} \frac{\partial S_i(t)}{\partial p_l}$$

Ingalls & Sauro, JTB, 2003



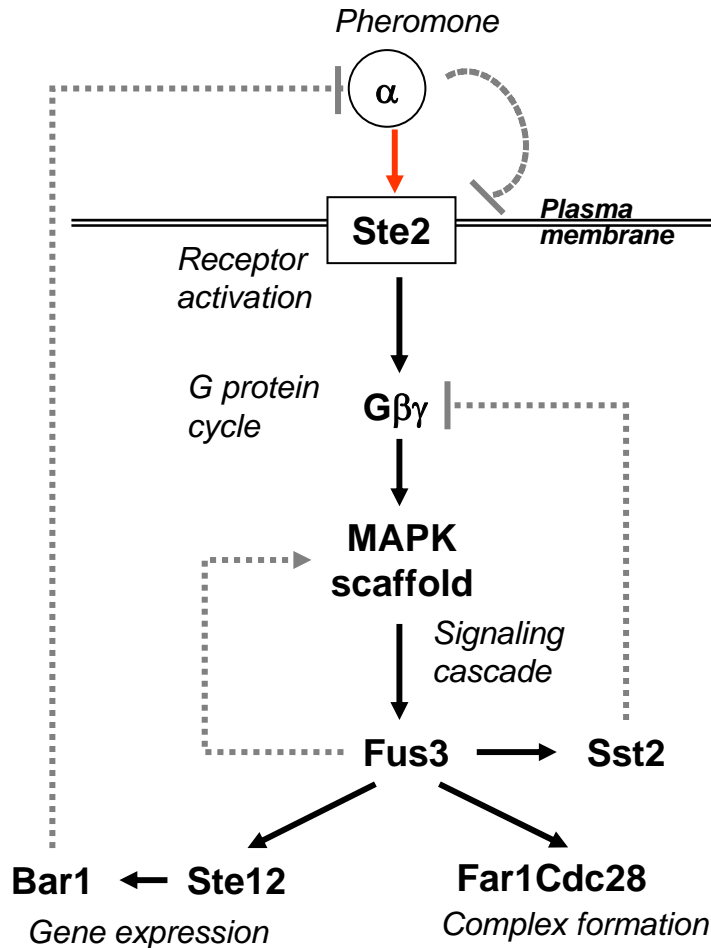


Signaling Pathways in Baker's Yeast

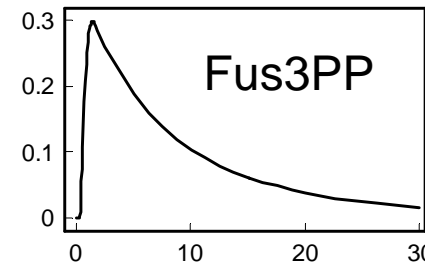




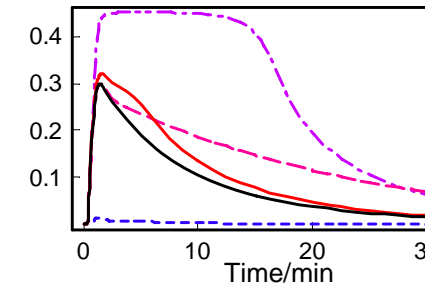
Pheromone Response



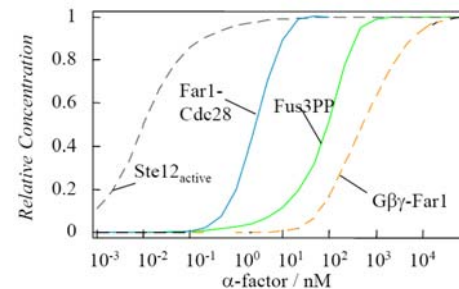
Simulation results:



Wild type



Mutants

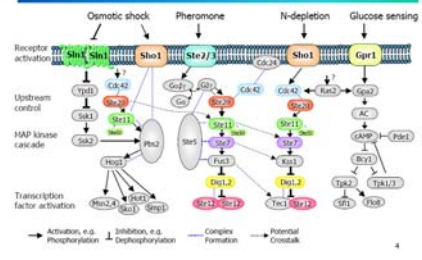


Graded response to dosage of α factor

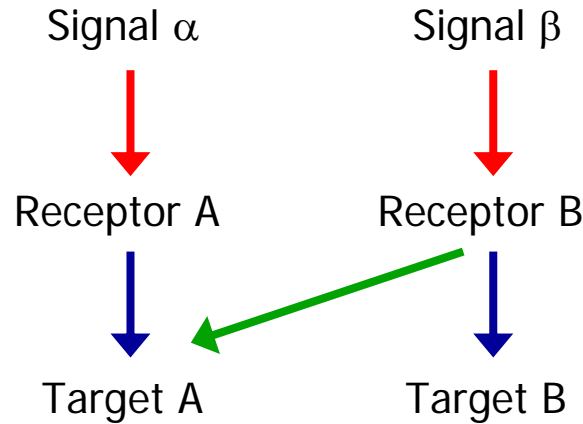
- *Transcription*
- *Cell cycle*
- *Mating projection*



Signaling Pathways in Baker's Yeast



Integration assessment

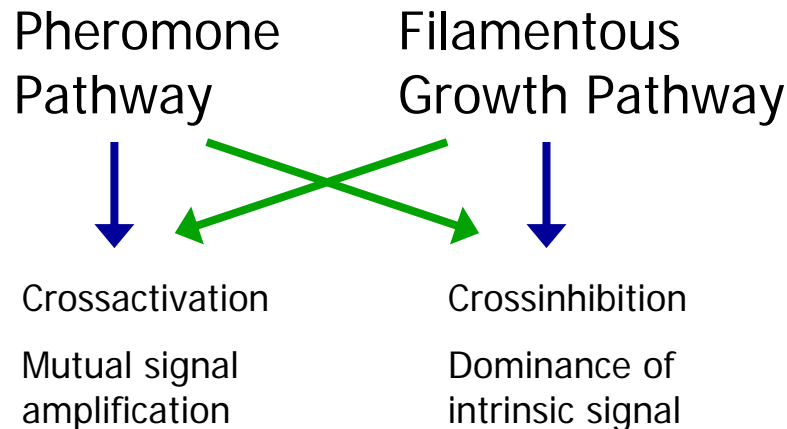


Measures of crosstalk

$$C = \frac{X(B)}{X(A)} \quad S_i = \frac{X(A)}{X(A,B)}$$

$$S_e = \frac{X(B)}{X(A,B)}$$

X – function of amplitude, timing or integral of response

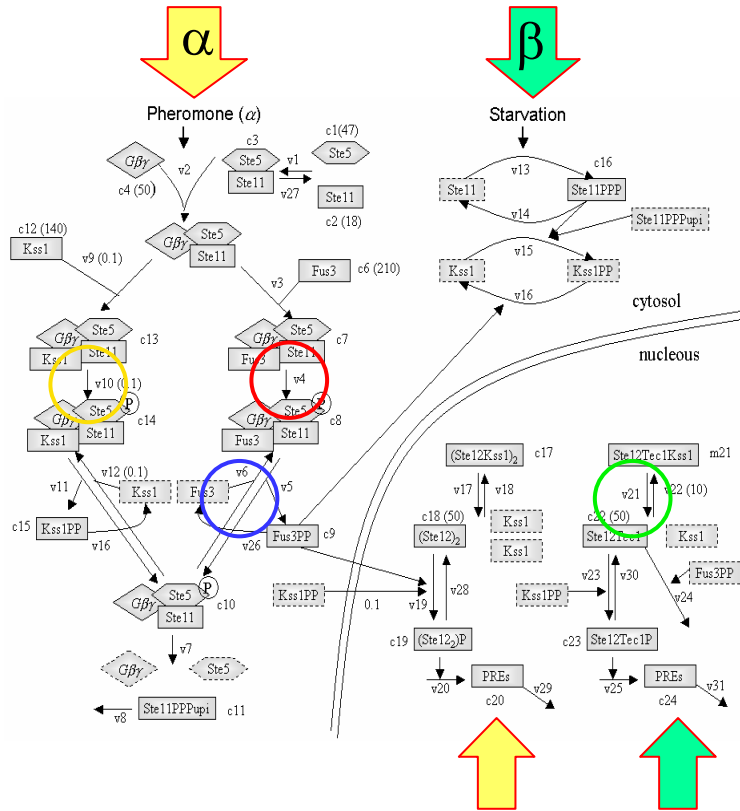


	$S_e > 1$	$S_e < 1$
$S_i > 1$	Mutual signal inhibition	Dominance of intrinsic signal
$S_i < 1$	Dominance of extrinsic signal	Mutual signal amplification

Schaber, Kofahl, Kowald & Klipp, 2006, FEBS J.



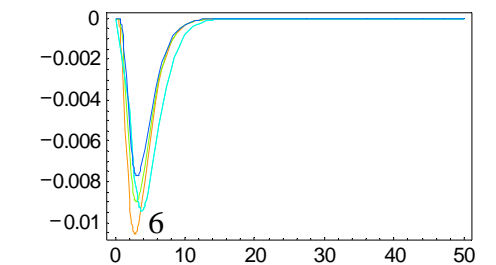
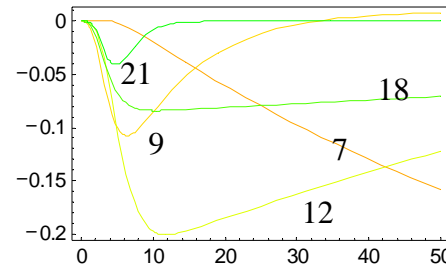
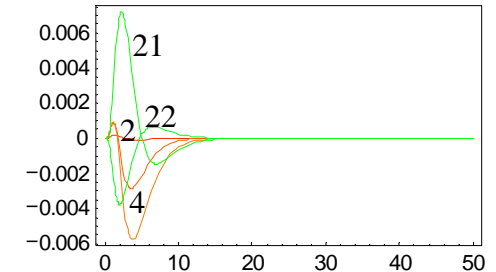
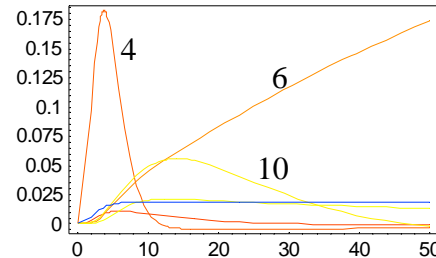
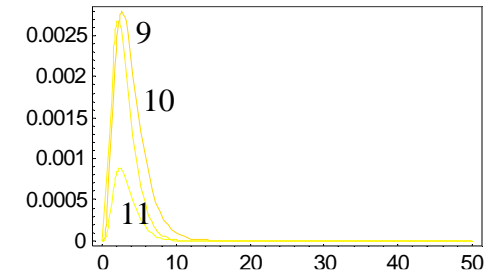
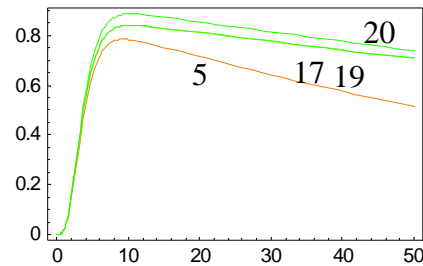
Integration of Signaling Pathways



Response coefficients of

PREs

FREs



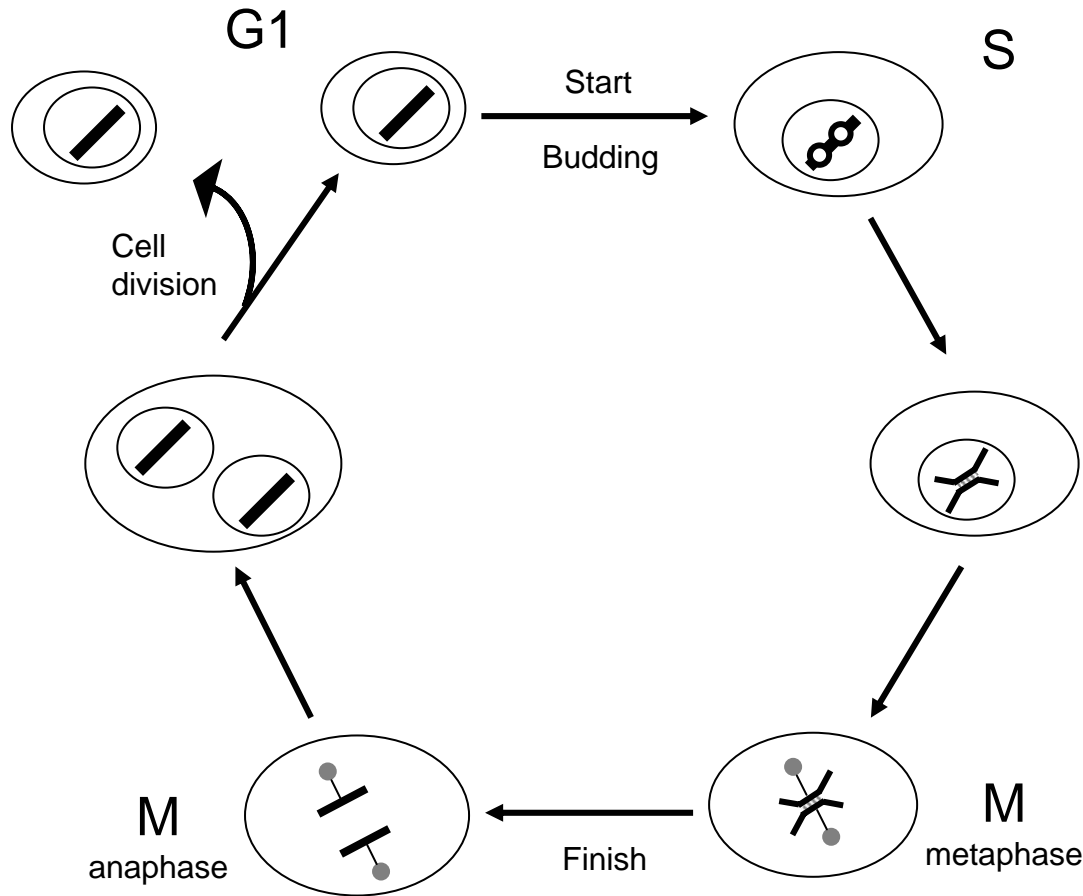
Time/min

Time/min

- 4 -Fus3 phosphorylation in MAPKcascade
- 6 -repeated Fus3 phosphorylation
- 10-Kss1 phosphorylation in MAPKcascade
- 21-Kss1 release from Ste12Tec1 complex



Yeast Cell Cycle



Cyclins *active*

Cln1-3, Clb5,6

Cyclin Dependent
Kinases (CDK)

Cdk1 (Cdc28)

Cyclin Dependent
Kinase Inhibitors (CKI)

Far1, Sic1 *inactive*

Transcriptional
Activators ←

SBF (Swi4-Swi6)
MBF (Mbp1-Swi6)

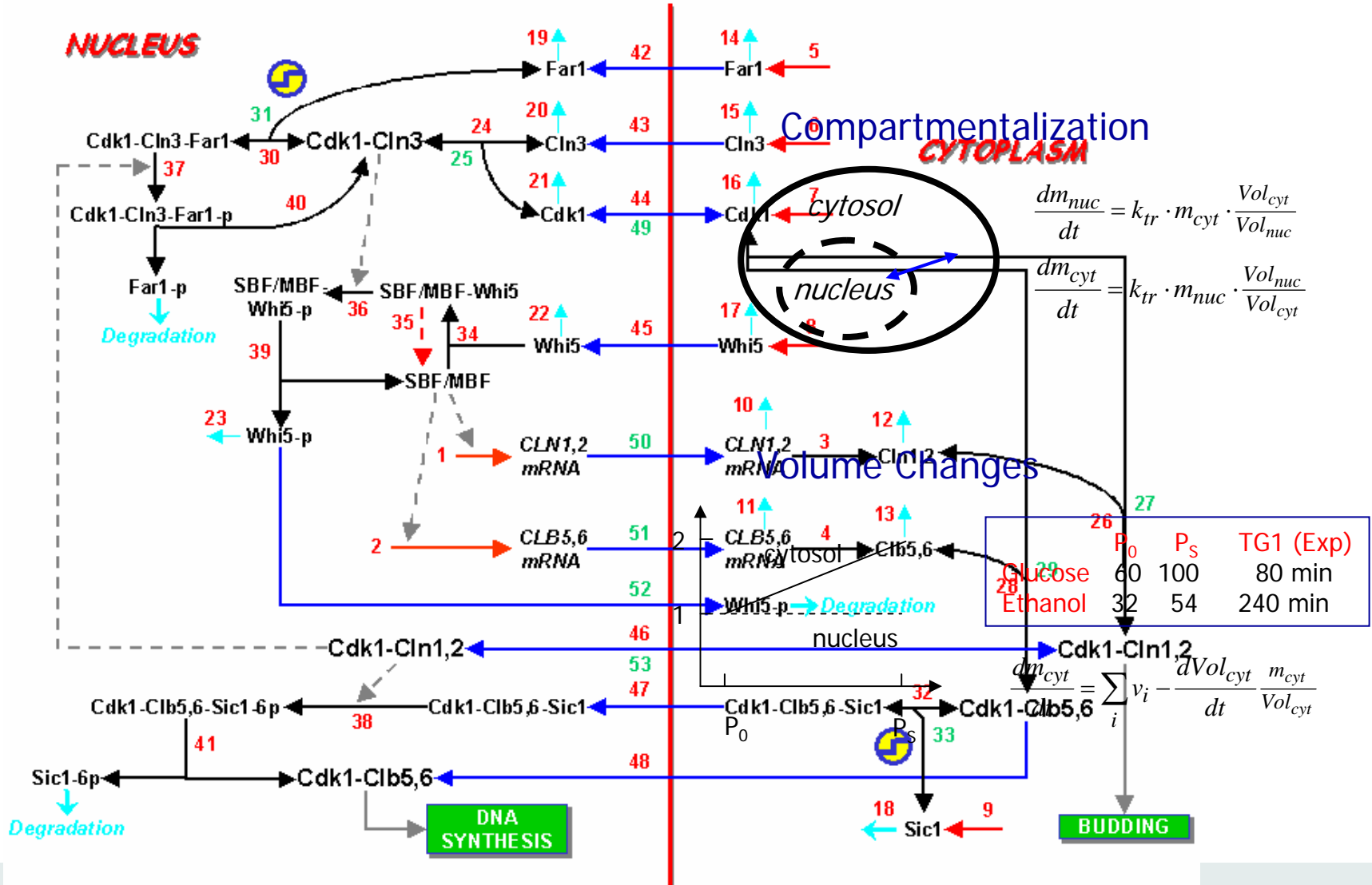
Transcriptional
Repressor ←

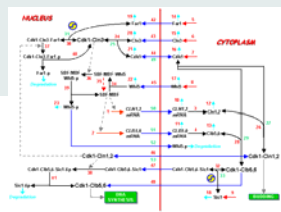
Whi5

In collaboration with Alberghina lab, University Milano-Bicocca



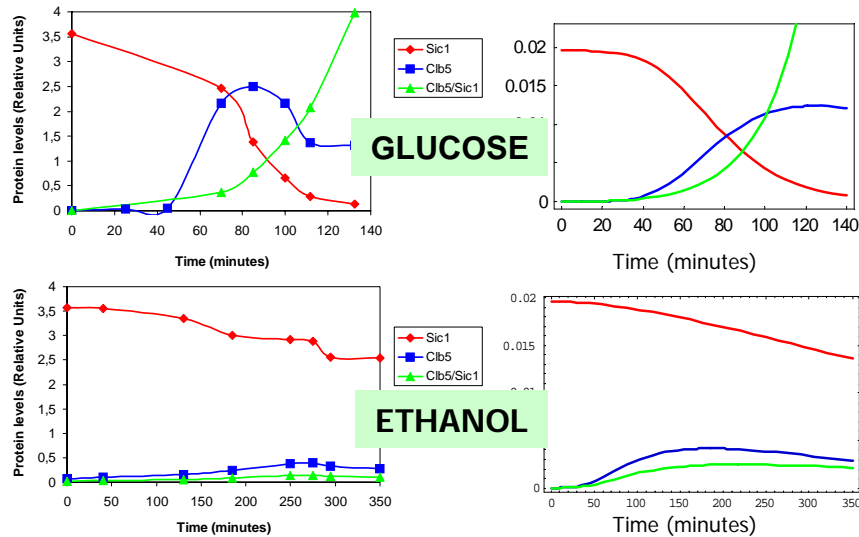
Network controlling the G1 to S transition





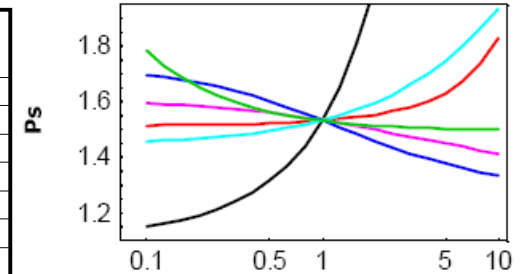
G1 to S transition: Model results

Reproduction of experiments



Explanation of critical cell size

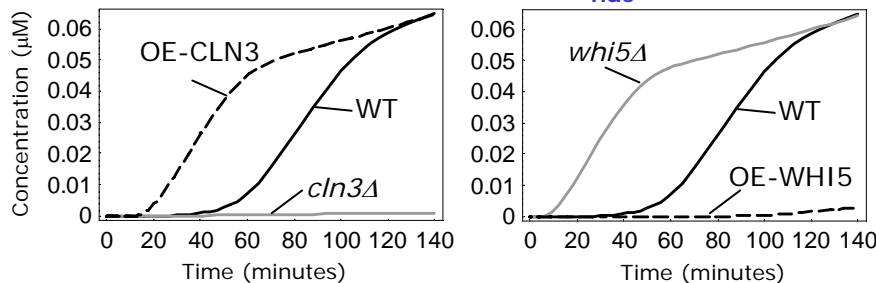
Relevant genotype	Estimated P_s
WT GLUCOSE	1.54
<i>cln3Δ</i>	-
GAL-CLN3	1.26
<i>far1Δ</i>	1.44
GAL-FAR1	-
<i>whi5Δ</i>	1.20
GAL-WHI5	3.31
<i>sic1Δ</i>	6.57
GAL-SIC1	1.50
WT ETHANOL	1.20



Growth rate
 Far1 production
 Cln3 production
 Far1 initial concentration
 Cln3 initial concentration
 Binding value Sic1/Cdk1-Clb5,6_{cyt}

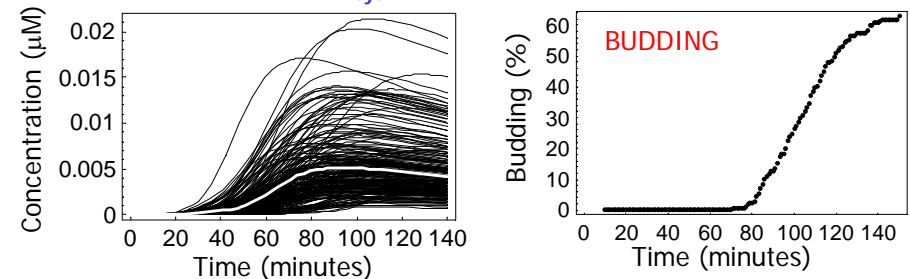
Prediction of mutant phenotypes

Cdk1/Clb5,6_{nuc}



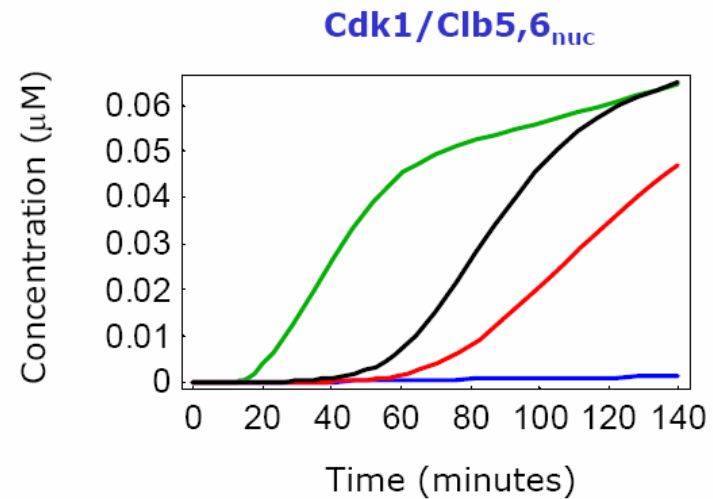
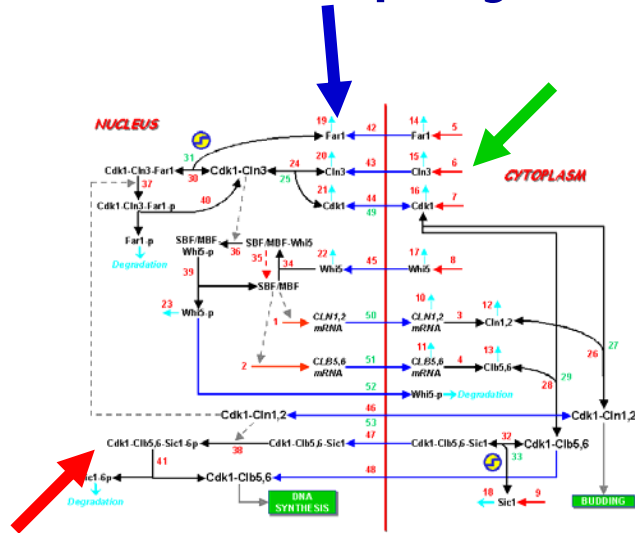
Population effects

Cdk1/Cln1,2_{cyt}

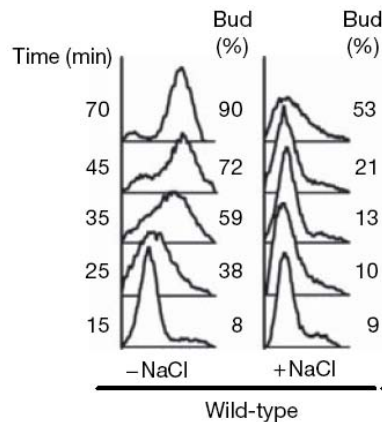




Interplay with signaling pathways



- Wildtype, no stress
- Pheromone pathway
- cAMP/PKA pathway
- HOG pathway/osmotic stress

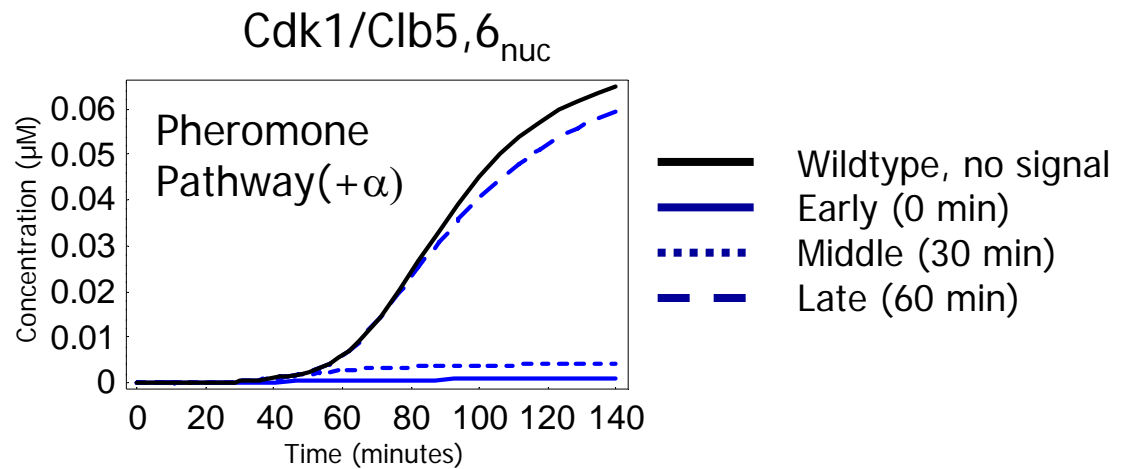
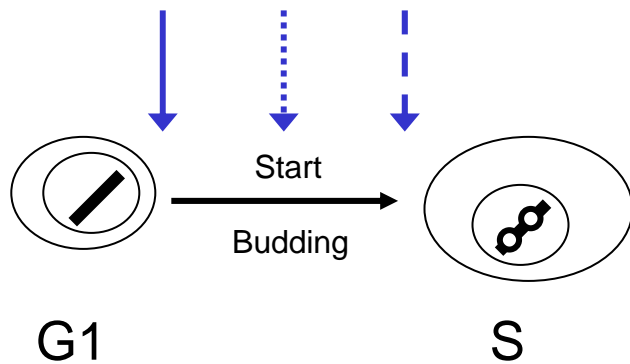


→ Delay or put forward G1/S transition

Escoté et al., Nature Cell Biol, 2004

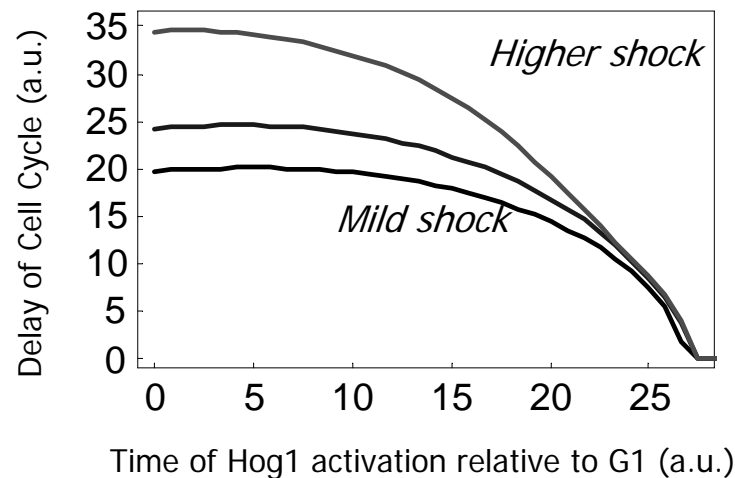
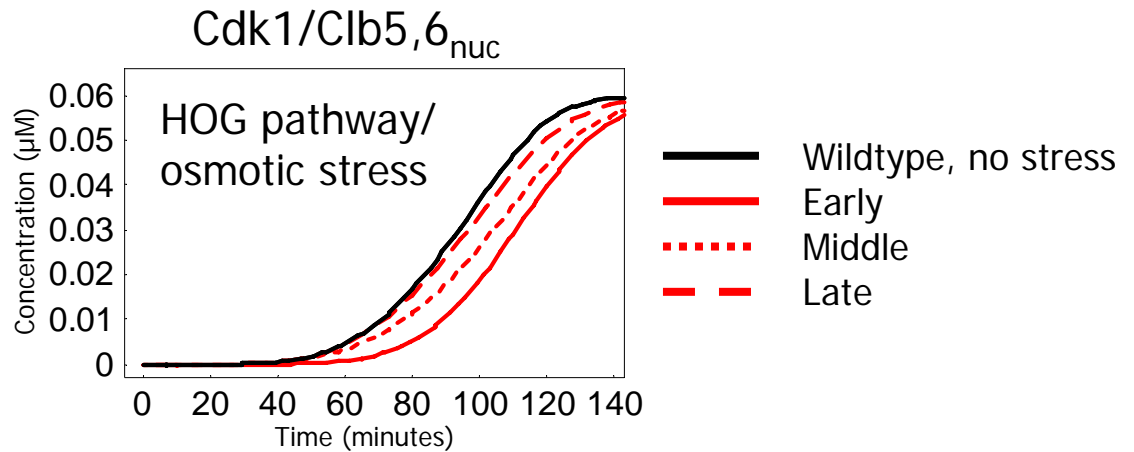
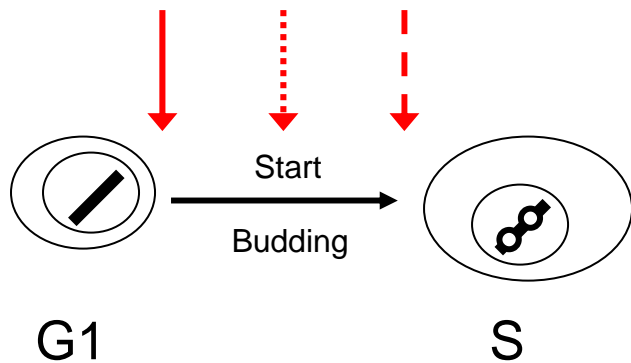


Timing of Signal Activation Determines Effect on Cell Cycle Progression





Timing of Signal Activation Determines Effect on Cell Cycle Progression



In collaboration with
Francesc Posas, Barcelona



Tool Development

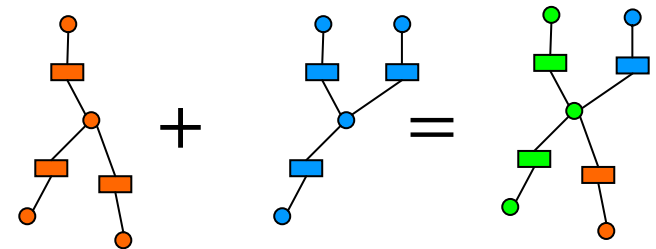
SBMLmerge Tool for model integration based on SBML representation

SBMLannotate → compounds and reactions

SBMLcheck → for consistency and SBML compatibility

SBMLmerge → merges valid models

SBML2dot → graphical output



<http://sysbio.molgen.mpg.de/sbmlmerge/>

Schulz, Uhlendorf, Liebermeister & Klipp, 2006, Genome Informatics

SBML-PET Tool for parameter estimation based on SBML representation

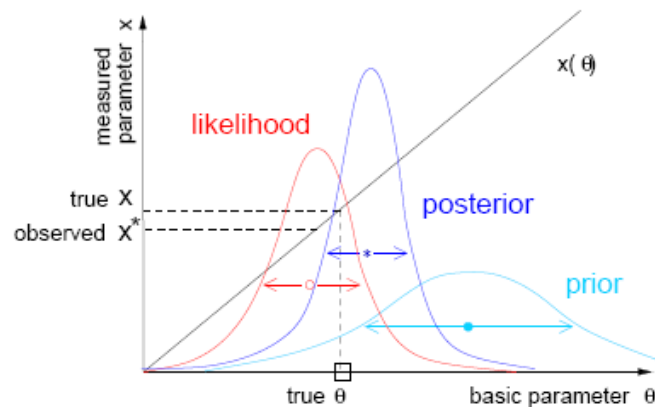
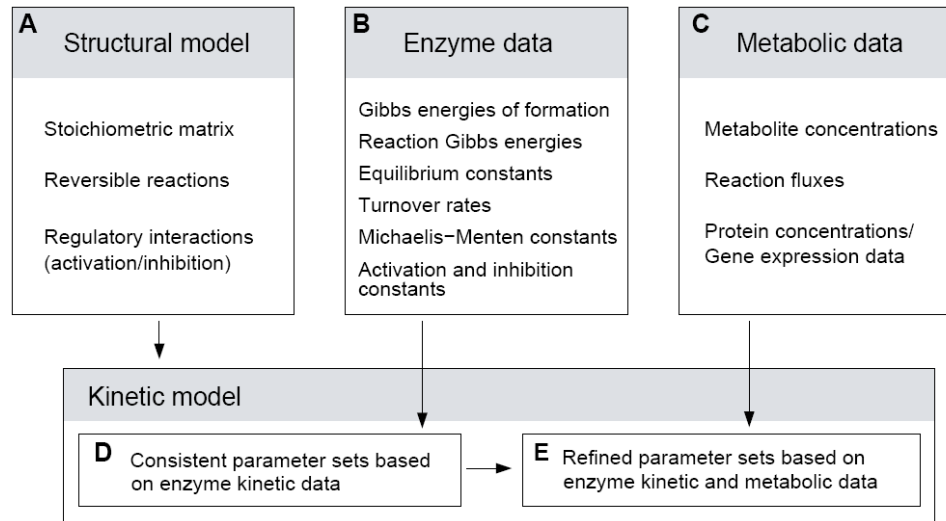
- SBML import and export
- Various types of data
- Various mathematical expressions
- Discontinuous state changes
- Supports Events

<http://sysbio.molgen.mpg.de/SBML-PET/>

Zi & Klipp, 2006, Bioinformatics



Data Integration



Assignment of Kinetics

Reaction network
+ thermodynamic info
+ kinetic info from DB
+ Convenience kinetics

Bayesian estimation

1. Parameter distribution

+ metabolomic data
+ genomic data

Bayesian estimation

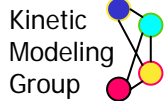
2. Parameter distribution

Liebermeister & Klipp, submitted



Conclusions

- Mathematical models of cellular processes allow for a testable representation of experimental knowledge.
- Models allow integration of diverse data and information.
- Modeling reveals regulatory properties of cellular network.
- The temporal organization of cellular events is critical to understand stress response – besides network structure and parameter values.



Acknowledgements

Wolfram Liebermeister

Jörg Schaber **BS01**

Simon Borger

René Hoffmann

Bente Kofahl

Sebastian Schmeier

Judith Wodke

Anselm Helbig

Marija Cvijovic

Zhike Zi

Christian Waltermann

Jannis Uhlendorf

Marvin Schulz

Axel Kowald

MPI Molecular Genetics

Roland Krüger

Reinhart Heinrich

Humboldt University Berlin

Stefan Hohmann

Bodil Nordlander

Per Sunnerhagen

Thomas Nyström

Göteborg University

Peter Gennemark

Chalmers University Göteborg

Lilia Alberghina

Matteo Barberis **BY14**

Romilde Manzoni

University Milano Bicocca

Matthias Peter

Fabian Rudolf

Serge Peletier



ETH Zürich



Francesc Posas

Miguel-Angel Andrade Nadal

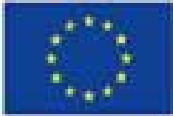

U Pompeu Fabra Barcelona


Funding is appreciated from


GEFÖRDERT VOM
 Bundesministerium für Bildung und Forschung → 

 Deutsche Forschungsgemeinschaft DFG → 


DAAD → PPP Schweden


 → 

→ 

→  Marie Curie Actions
Human Resources and Mobility Activity

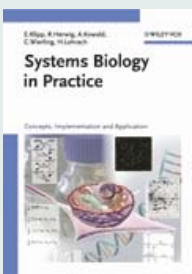
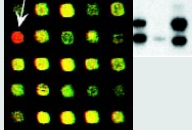
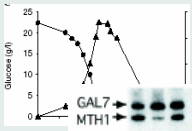
Early Stage Training
Systems Biology

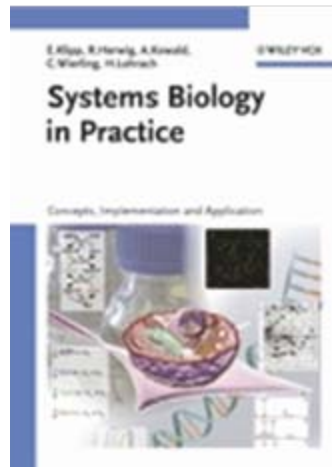
→ ENFIN 

→  The Yeast Systems Biology Network YSBN



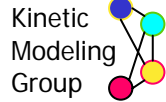
$$\begin{aligned} dX_1/dt &= v_0 - v_1 \\ dX_2/dt &= 2v_1 - v_2 - v_9 \\ dX_3/dt &= v_2 - v_3 + v_4 - v_5 \\ dX_4/dt &= v_3 - v_4 \\ dNADH/dt &= v_2 - v_5 + v_4 + 4v_5 - v_6 \\ dATP/dt &= -2v_1 + 2v_2 + 3v_6 - v_7 \end{aligned}$$





Textbook on Systems Biology

By Klipp E, Herwig R, Kowald A, Wierling C, Lehrach H
WILEY-VCH, 2005



Acknowledgements



http://www.molgen.mpg.de/~ag_klipp



$$\begin{aligned} dX_1/dt &= v_0 - v_1 \\ dX_2/dt &= 2v_1 - v_2 - v_9 \\ dX_3/dt &= v_2 - v_3 + v_4 - v_5 \\ dX_4/dt &= v_3 - v_4 \\ dNADH/dt &= v_2 - v_5 + v_4 + 4v_5 - v_6 \\ dATP/dt &= -2v_1 + 2v_2 + 3v_6 - v_7 \end{aligned}$$

